

Newsletter of the Workgroup Pineapple, International Society for Horticultural Science Issue No. 23, July, 2016

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# **Pineapple Working Group News**

#### From the Editor

Dear Colleagues:

# IX International Pineapple Symposium will not be held in Costa Rica

See below details about the cancelation from Workgroup Pineapple Chair Domingo Haroldo Reinhardt.

#### **Thanks**

I wish to extend my thanks to all who continue to contribute to and support Pineapple News. As a result of the many interesting contributions provided by "Pineapple People" throughout the tropics over many years Pineapple News celebrates its 23<sup>rd</sup> year of helping to keep readers informed about the many issues related to pineapple. With the continued support of all those "People", we will meet again in about a year.

## Pineapple taxonomy & cultivar naming

The taxonomy of pineapple was greatly simplified in 2003 (Coppens d'Eeckenbrugge and Leal, 2003) and a recent tweak has made it possible for Coppens d'Eeckenbrugge to declare below in **News from France** that it is *The Last Revision of Pineapple Nomenclature*. While the taxonomy of pineapple appears to have been clarified and solidified, the naming of pineapple cultivars remains a serious problem. Many of the problems are encountered in research papers on pineapple published in English by authors who identify experimental material by local or trade names rather than by the cultivar name.

Technically a cultivar has only one correct name and it is the cultivar name that joins new research results to the body of literature that includes all relevant publications about a pineapple clone or cultivar. I don't have the expertise to declare this to be a unique problem with pineapple. However, it is certainly unique when compared with temperate fruit crops and I speculate that the problem with pineapple is partly related to its discovery and its wide distribution in the early years after discovery. The three main cultivars, 'Queen', 'Smooth Cayenne' and 'Spanish', lost whatever names they might have had when early explorers took them from their country of origin. Using 'Smooth Cayenne' as an example, it acquired new names as it was dispersed throughout the tropics and brought to Europe and Great Britian (Collins, 1950). It was first named Cayenne Lisse in France, became 'Smooth Cayenne' (the English translation) in Great Britian, Kew when the English took it to India and Sarawak when it was taken to Malaysia. More recently papers published by researchers from Thailand show it has acquired new names in that country. In addition, there are multiple named clones for each of the three cultivars mentioned above that are often treated as cultivars even though no formal description was published to support the new name

The first article below was written to identify some of the recent problems with cultivar naming and provide recommendations that, if followed, will provide a strategy to reduce the problem of misnamed cultivars. The proper naming of a cultivar allows research on it to be linked to all other research on the cultivar while incorrect naming relegates such research to some obscure corner where it may or may not be found.

The second article below highlights what has been done to date to publish the names of new pineapple cultivars, introduces a plan to involve the ISHS Workgroup Pineapple in supporting a cultivar naming effort and introduces a list, by country, of pineapple cultivar names and at least some of their synonyms. It is hoped that Workgroup Pineapple members will support these two efforts to assure that pineapple cultivar descriptions are publicly available ant that there is easy access to synonym names in hopes that both efforts will insure that cultivars are properly named in research papers.

### Pineapple News also now available without a log in at Univ. of Hawaii Scholar Space

https://scholarspace.manoa.hawaii.edu/handle/10125/41067

Duane P. Bartholomew

#### Reference

Collins, J.L., 1951. Notes on the origin, history, and genetic nature of the Cayenne pineapple. Pacific Sci 5:3-17.

# **Proceedings of the 8<sup>th</sup> International Pineapple Symposium**

Acta Horticulturae 1111 (<a href="http://www.ishs.org/pineapple">http://www.ishs.org/pineapple</a>) lists the papers, including abstracts, that were presented at the VIII International Pineapple Symposium. Because the 8<sup>th</sup> International Pineapple Symposium was part of the XXIX Horticultural Congress held in Brisbane, Australia in 2015, the Acta volume that contains all of the papers on pineapple that were submitted for the proceedings also contains the papers on guava and mango. The above page also provides a link to all back issues of Pineapple News under the heading THIS WORKGROUPS PAGES.

# **Proper Naming of Pineapple Cultivars: Problems and Recommendations for Improvement**

D.P. Bartholomew

As a referee of papers on pineapple submitted for publication in technical journals and as an editor of papers and proceedings of the various international pineapple symposiums I have become concerned about the lack of standardization in the naming of pineapple cultivars. Clones of a known cultivar and local names based on geographic location or to attract consumers are not cultivar names and should not be used in published research papers unless they are accompanied by the cultivar name.

Using the correction name of a cultivar connects new research to the body of published literature, which allows others interested in the cultivar to easily locate all of the published research on the cultivar. See below that proper identification of pineapple cultivars is also a concern of others.

Coppens d'Eeckenbrugge (2014). A widely accepted plant nomenclature, based on precise identifications and descriptions, is essential in defining properly the objects of our scientific and economic activities and making it possible to understand each other and compare results.

Zhou et al. (2015). However, the exchange of vegetative planting materials has also resulted in problems for conservators of pineapple germplasm because records and labels of the cultivars have not always followed the same naming conventions, and accessions have limited information about their correct identity. Therefore, homonyms and synonyms are common among the names of pineapple cultivars and that restricts the sharing of information and materials among pineapple researchers and hampers the use of pineapple germplasm in breeding. Another major challenge for pineapple cultivar identification is that the protracted vegetative propagation has led to the accumulation of somatic mutations. Some mutations caused noticeable phenotypic effects and created intra-cultivar variation, which became the target of clonal selection. While these selected mutants are important in horticultural production, it is necessary to identify them so that breeders and genebank curators can efficiently conserve and use these genetic materials.

In some cases the solution to the problems are obvious. In all cases using the correct cultivar name is the responsibility of the author(s). The proliferation of scientific journals in the current era of rapid review and publication on the World Wide Web makes it nearly impossible for reviewers and editors to focus on the proper identification of a cultivar. Improvement can only come if there is an organized effort to bring order out of the chaos of names.

#### **Examples of cultivar identification problems**

Use of local or clone names to identify a cultivar:

• 'Pérola', 'Queen' and 'Smooth Cayenne' have been given various synonyms in multiple countries. The greatest proliferation of such names is relatively recent and mainly in Asia and Africa.

- Sugarloaf and assorted variations of it (Sugar Loaf, sugarloaf), probably in most cases 'Pérola', have been used in the Caribbean for generations. Collins (1960) stated that Sugar Loaf does not refer to a specific variety and Johnson (1935) lists eight varieties that have Sugar Loaf as a synonym. Sugar Loaf, or its French translation, Pain de Sucre, is used in research papers by authors from Benin, Ghana and Malaysia.
- 'Smooth Cayenne' (first mentioned as 'Cayenne lisse' in 1820; as Smooth Cayenne in 1886), also known as 'Kew' in India (first published reference in 1942) and 'Sarawak' (first published reference in 1937) in Malaysia, also has several named clones that are treated by some authors as cultivars. In publications from Thailand synonyms treated as cultivars include the local names Lakata, Pattavia, perhaps Bathavia, Singkapropattavia, Petburi No. 2 and Nanglae (said to be regional sub-variety of Pattavia. In some cases what are listed by authors as cultivars may be varied spellings of the same Thai name.

## Inconsistent naming of cultivars

- See above regarding inconsistent naming of what probably are 'Smooth Cayenne' clones in Thailand.
- Even minor name changes can be confusing and limit access to publications. For example, Pérola and Pearl (the latter the English translation of the name) and Fantástico and Fantastic have been used in papers published by authors from Brazil. Fortunately, database searces for Pérola and Perola return the same 217 references while that isn't the case for Pérola and Pearl.
- 'Victoria' and 'Queen Victoria', not to be confused with 'Vitória' (also called 'Victoria' in a paper from Brazil), a cultivar developed by Embrapa (Brazil), from a cross between 'Primavera' and 'Smooth Cayenne' (Bartholomew et al., 2010), are used interchangeably in references originating in Mauritius, Reunion and South Africa. Based on the publication history, they are the same 'Queen' clone, first referred to as 'Queen Victoria' in Mauritius in 1972, referred to as Victoria there in 2001, named Victoria and described in Reunion in 1976 and varyingly referred to thereafter as 'Victoria' and 'Queen Victoria' in Reunion and South Africa. To add to the confusion, a Queen Victoria RE43 was reported in Côte d'Ivoire in a 2013 publication.

#### Naming cultivars developed in countries that do not use the Roman alphabet, e.g., Taiwan

• Tainung cultivars. These hybrids were developed in Taiwan. They have been identified as TN followed by the number and 'Tainon No. etc.' by Taiwanese authors and as 'Tainong No. etc.' in papers by Chinese authors. Minor variations in spelling of the Tainung cultivars can cause confusion and inconsistent search results. The problem is in the translation from the Chinese because when using <a href="https://translate.google.com/">https://translate.google.com/</a>, Tainung and Tainong result in the same character (logogram) set.

#### Typographical error/misspelling

- Moris ('Queen') In a few instances the clone is misspelled as Morris.
- 'Josapine' (a name remembering the parents of this Malaysian hybrid: 'JOhor' x 'SArawak'). Josephine was used in an abstract and in a recent paper.

#### General recommendations:

- 1. Papers reporting the results of research on pineapple in a language using the Roman alphabet that do not correctly identify the cultivar must be returned to the author(s) for correction.
- 2. Authors should use the earliest correct name of a cultivar because it takes precedence and avoids confusion. A complete clone and cultivar list is being prepared as a guide (see *A New Source of Pineapple Cultivar Names and Descriptions* below).
- 3. For countries that use the Roman alphabet, the correct cultivar name is the name in the language of the country, e.g., Pérola, not Pearl.
- 4. Standardize the translation of cultivars developed in the Asian countries that use characters (logograms) instead of the Roman alphabet.
- 5. When referring to a local name or a clone of a known cultivar, authors should follow the practice described by Coppens d'Eeckenbrugge et al. (1997), which is to provide the local or clone name along with the cultivar name, for example, Victoria ('Queen'), Sugar Loaf ('Pérola'), Champaka F-153 ('Smooth

Cayenne'). No "cv" prefix or single quotation marks are used around the clone or local names because they are not cultivars.

**Acknowledgement**: Thanks are due to G. Coppens d'Eeckenbrugge and G. Sanewski for their helpful editing and suggestions.

Coppens d'Eeckenbrugge, G. 2014. Pineapple taxonomy: Species, botanical varieties and cultivars, and their importance in understanding and managing pineapple diversity. Pineapple News No. 21, 34-37. <a href="http://www.ishs.org/pineapple/pineapple-newsletters">http://www.ishs.org/pineapple/pineapple-newsletters</a>

Coppens d'Eeckenbrugge, G., Leal, F., and Duval, M.F. (1997) Germplasm resources of pineapple. Horticultural Reviews 21, 133-175.

Collins, J.L., 1960. The Pineapple: Botany, Cultivation and Utilization. Interscience Publishers Inc., New York. Johnson, M.O., 1935. The Pineapple. Paradise of the Pacific Press, Honolulu.

Zhou, L., Matsumoto, T., Tan, H., Meinhardt, L.W., Mischke, S., Wang, B., and Zhang, D., 2015. Developing single nucleotide polymorphism markers for the identification of pineapple (Ananas comosus) germplasm. Horticulture Research 2:(25 November 2015).

# **New Sources for Pineapple Cultivar Names and Descriptions: Contributions from "Pineapple People" Invited**

D. P. Bartholomew, G. Coppens d'Eeckenbrugge and G. M. Sanewski

The American Society for Horticulture Science (ASHS) has quite a long history of publishing cultivar names and descriptions of fruits and nuts in the journal HortScience (Cummins, 1994). Brooks and Olmo (1997) published a compendium of fruit and nut varieties that included the descriptions of the pineapple cultivars CO-2, Smooth Cayenne and Spanish Jewel. The description of Smooth Cayenne included Pineapple Research Institute of Hawaii hybrids '53-116' and '59-656' that should have been listed under their own headings.

No recent compendiums of fruit and nut varieties have appeared since 1997. However, HortScience continues to publish Fruit and Nut variety descriptions every two years. Pineapple has not been well represented in the HortScience Register until recently. In 2009 a list of 17 cultivar descriptions were included in Register of New Fruit and Nut Cultivar List 45 (Bartholomew et al., 2010; see list at

http://hortsci.ashspublications.org/content/45/5/716.short). No pineapple cultivar descriptions were contributed to Lists 46 and 47 but an additional 10 cultivar descriptions were contributed to Register of New Fruit and Nut Cultivar List 48 (Bartholomew et al., 2016; see list at

http://hortsci.ashspublications.org/content/51/6/620.full.pdf+html). While the HortScience lists are freely available, we are hoping to develop a complete alphabetical list, including photographs, which would provide a more efficient way to access pineapple cultivar descriptions.

#### **Searchable List of Pineapple Cultivar Descriptions**

If the "Pineapple People" (Butcher and Gouda, 2014), i.e. the Pineapple Working Group of ISHS (<a href="http://www.ishs.org/pineapple">http://www.ishs.org/pineapple</a>), can promote and sustain an effort to publish cultivar information in HortScience, we should soon have a relatively complete list of pineapple cultivars of some commercial importance on national and international markets. Our objective is to create an on-line searchable list of the commercially important cultivars and clones as well as those that are considered to be valuable parents in a pineapple breeding program.

### Factsheet to Guide Preparation of Pineapple Cultivar Descriptions

We also plan to develop a cultivar factsheet that can be used to submit information about new cultivars that should be added to the HortScience Register of Fruits and Nuts. Once the factsheet has been developed it will share with readers by email to the Pineapple News mailing list.

# **Inventory of Diverse Pre-Columbian Pineapple Materials**

We also propose to find collaborators who will be willing to help develop an inventory of the highly diverse pre-Columbian materials, marketed at small scales in tropical Latin America. This is expected to require considerable time and be difficult as they are most often mentioned only in the grey literature so accurate descriptions likely would need to be developed. The Factsheet referred to above will be developed with this objective in mind.

## Development of an On-line List, by Country, of Pineapple Cultivars.

In support of the above efforts, D.P. Bartholomew searched through the pineapple reference database, now containing over 10,000 references, by country and cultivar and collected a large number of cultivar and synonym names. The names, by country, have been accumulated in a spreadsheet that was reviewed by the coauthors and their additions and corrections have been included in the list. The list can be viewed at: <a href="https://docs.google.com/spreadsheets/d/1NDr9v3FSZLP8W3m9rYhScErrxed8vEc7f8meDyLXEmg/pubhtml">https://docs.google.com/spreadsheets/d/1NDr9v3FSZLP8W3m9rYhScErrxed8vEc7f8meDyLXEmg/pubhtml</a> and contains the following information:

- 1. Names of cultivars and hybrids found in the published literature from that country.
- 2. Parents of hybrids where known.
- 3. Synonyms, clones, local names used for the cultivar in the country.
- 4. The earliest reference that named the cultivar or contained information about it.

#### What Can Readers Do

- Help keep the published list of pineapple cultivars current. G. M. Sanewski
   (garth.sanewski@daff.qld.gov.au) has agreed to lead and coordinate this effort. He will develop a
   Factsheet to be used in developing pineapple cultivar descriptions that conform to the guidelines of
   HortScience. If you or someone you know is developing new pineapple cultivars to please contact G. M.
   Sanewski.
- 2. Help develop an inventory of Pre-Columbian pineapple materials currently being grown in Central and South America. Please contact G. Coppens d'Eeckenbrugge (geo.coppens@cirad.fr) if you can help with this project.
- 3. Help maintain a current by-country database of pineapple cultivars and their synonyms. The final objective for the list would be to have it contain the names of all cultivars that are considered to be important commercially or as parents in breeding programs. Please contact D.P Bartholomew (duaneb@hawaii.edu) with additions and corrections to the spreadsheet.

#### References

Bartholomew, D.P., G. Coppens d'Eeckenbrugge, G., and C.C. Chen. 2010. Register of New Fruit and Nut Cultivar List 45 (Pineapple, pp. 740-742). HortScience 45:716-756.

Bartholomew, D.P., G.M. Sanewski and C.C. Chen. 2016. Register of New Fruit and Nut Cultivar List 48 (Pineapple, pp. 640-641). HortScience 51: 620-652.

Butcher, E. and D. Gouda. 2014. Most Ananas are cultivars. Pineapple News No. 21, 9-11.

http://www.ishs.org/pineapple/pineapple-newsletters

Brooks, R.M. and Olomo, H.P. 1997. Pineapple, p. 580. The Brooks and Olmo register of fruit & nut varieties. ASHS Press. 743 pages.

Coppens d'Eeckenbrugge, G. 2014. Pineapple taxonomy: Species, botanical varieties and cultivars, and their importance in understanding and managing pineapple diversity. Pineapple News No. 21, 34-37. http://www.ishs.org/pineapple/pineapple-newsletters

Cummins, J.N. 1994. New System for Fruit and Nut Variety Naming and Registration. Fruit Var J. (APS) 48:114-116. <a href="http://www.pubhort.org/aps/48/v48\_n2\_a10.htm">http://www.pubhort.org/aps/48/v48\_n2\_a10.htm</a>.

# **News from Brazil**

# IX International Pineapple Symposium – Plans for Costa Rica Canceled

Domingo Haroldo Reinhardt, Chair, ISHS Workgroup Pineapple

Contacts done by the ISHS Workgroup Pineapple (WGP; <a href="http://www.ishs.org/pineapple">http://www.ishs.org/pineapple</a>) after the last IPS held in Brisbane, Australia, in late 2014 led to the identification of the convener (Dr. Luis Pocasangre) and the institution (University Earth) to coordinate the organization of the IX IPS in Costa Rica, one of the largest producers and the leading exporter of fresh pineapple in the world. Until very recently everything seemed to be all right and the agreement with ISHS was underway. However, on July 19 WGP received a letter from Dr. Pocasangre communicating that he and Earth University are canceling their intention to organize the event in Costa Rica due to the risk of introduction of the Tropical race 4 strain of *Fusarium oxysporum f. sp. cubense* (FOC), pathogen of Panamá wilt, which is known to be the most devastating disease of banana plantations in Asia, has recently shown up in Africa, but has not yet been detected in the Americas. Pineapple production in Costa Rica is found in the same region where the main banana production occurs and the big players are mostly the same companies.

PWG has started contacts to define a new site for the event. The preference would be for a country in the American continent, but international concerns about FOC may favor the event location to be focused in a country where that disease is already present or where banana is not an important industry.

Any news on this will be communicated to the WPG and Pineapple News mailing lists. Any useful contribution is welcome and should be sent to Prof. Duane Bartholomew (<u>duaneb@hawaii.edu</u>) and Domingo Haroldo Reinhardt (<u>domingo.reinhardt@embrapa.br</u>).

# VI Brazilian Pineapple Symposium

Stella de Castro Santos Machado, Geraldo Chaser Tavares, Aristoteles Pires de Matos, Domingo Haroldo Reinhardt

The VI Brazilian Pineapple Symposium was held under the theme "strengthening the pineapple industry with social and environmental responsibility", in Conceição do Araguaia, Pará, North Brazil, from 11 to 13 November 2015. That is a region where more than 450 thousand metric tons of pineapples are being produced every year by more than a thousand growers mainly under conditions of family agriculture.

There were 700 participants, mostly farmers, researchers, rural extension specialists, teachers and students, all with great interest in recycling and updating their knowledge on pineapple. The participants came from 74 pineapple producing municipalities of eight Brazilian states (Acre, Pará and Tocantins in the North, Bahia, Maranhão and Paraíba in the Northeast, Mato Grosso in the Center-West and São Paulo in the Southeast).

The main themes addressed at the symposium were mechanization, integrated management of pests, climatic changes and their impacts, organic farming, harvest and post-harvest handling. Several interesting round table debates were carried out focusing on the improvement of public policies in support of the pineapple industry, from production to marketing. On the evenings participants and local inhabitants could taste typical regional music and pineapple-based food.

The event was closed with a Field Day carried out at 'Pérola' pineapple fields of a typical small farmer's settlement with demonstrations on sprinkler irrigation systems, mechanized planting and pest control procedures with emphasis on fusariosis. The organizing committee formed by a partnership between Embrapa, Federal Institute of Pará (IFPA), Technical Assistance and Rural Extension Corporation of Pará (EMATER-PA), Agricultural Protection Agency of Pará (ADEPARÁ), the Brazilian Support Service for Micro and Small Enterprises (Sebrae), the State Department of Agriculture and Fisheries Development (SEDAP) and the Municipalities of Conceição do Araguaia and Floresta do Araguaia.

The next Brazilian Pineapple Symposium will be held in the state of Maranhão in 2016 or 2017.

# **Embrapa Held First Course on Cryopreservation of Plants with Emphasis on Pineapple**

Domingo Haroldo Reinhardt.

Thirteen participants from different parts of Brazil took part in the 1st Course on Cryopreservation of Plants held at Embrapa Cassava & Fruits in Cruz das Almas, Bahia, Brazil, from July 11 to July 15, 2016. The event was led by Fernanda Vidigal Duarte Souza, research scientist in tissue culture and genetic resources at Embrapa, and Maria lena González-Benito, professor at the Polytechnic University of Madri, Spain, one of the main international specialist on cryopreservation.

Vegetal cryopreservation is a long term conservation of plants based upon deep freezing conditions in liquid nitrogen at -196 °C. This course was focused on the application of this technique on seeds and plant tissues, especially on pineapple, banana and sugarcane, with a strong theoretical basis and large segments with hands-on practicing, a reason for the limited number of participants. In 2012, Fernanda Souza earned a post-doc period focused on this technique at the National Center for Genetic Resources and Preservation, Fort Collins, Colorado, USA, and is on the way to establish a cryobank for pineapple germplasm at Embrapa, a rather efficient alternative for long term conservation of valuable genotypes.

# Organic Pineapple Production System for the Region of Lençóis, Chapada Diamantina, Bahia, Brazil

Tullio Raphael Pereira de Pádua, Aristoteles Pires de Matos, Ronielli Cardoso Reis, Eliseth Souza Viana, Raul Castro Cariello Rosa, Francisco Alisson da Silva Xavier and Domingo Haroldo Reinhardt Embrapa Mandioca e Fruticultura, Cruz das Almas, Bahia, Brazil

In partnership with the company Bioenergia Orgânicos, Embrapa Mandioca e Fruticultura has been carrying out studies towards the establishment of organic production systems for several fruits, including pineapple. In the first stage, the focus has been on fertilization, planting density and weed control for the new 'BRS Imperial' pineapple and the traditional 'Pérola'. These are aspects related to the important role played by soil components in organic farming. There is always a need to improve the physical, chemical and biological properties of the soil to this environment, aiming to assure adequate plant and fruit development. In addition, planting density is a variable with strong impacts on yield, fruit size and fruit quality to meet the requirements for fresh fruit markets and industrial processing.

The studies have been carried out in the municipality of Lençóis, Chapada Diamantina, Bahia, a region with a sub-humid climate, a rainy season from November to April, when the temperature ranges from  $20~^{\circ}$ C to  $30~^{\circ}$ C, and a dry season from May to October with temperatures between  $17~^{\circ}$ C and  $26~^{\circ}$ C.

Five fertilization doses, 10, 20, 30, 40 and 50 t/ha, of cattle manure and organic Bokashi compost have been studied, as well as five planting densities (26,315; 31,250; 35,714; 47,620 and 51,282 plants/ha) corresponding to the spacings of 1.5 m x 0.4 m x 0.4 m; 1.2 m x 0.4 m x 0.4 m; 1.0 m x 0.4 m; 1.0

Before planting of pineapple cultivars and after correction of soil acidity by liming, soil preparation was done by sowing a cocktail of cover crops composed of a mixture of grasses (millet and sorghum) and legumes (velvet bean and jack bean) (Figure 1). Seeds were scattered by hand over the soil surface. As a complement of soil fertilization 250 kg/ha of natural phosphate was also applied. The shoot biomass produced by cover crops was mowed 80 days after sowing and kept on the soil surface for natural degradation (Figure 2). Fifteen days later the two pineapple cultivars were planted and 350 g of cattle manure and 250 g of powder-rock (calcosilicataded pyroxenite) were applied per plant.







Figure 1. Sowing and growth of soil cover crops (cocktail) in pre-planting of Pineapple. Lençóis, Bahia, Brazil. (Photos by Padua, T.R.P.)



Figure 2. Mowing of the cover crop biomass 80 days after sowing followed by the planting of pineapple cvs. BRS Imperial and Pérola. (Photos by Padua, T.R.P.)

Pineapples were harvested in August, during the dry season. Average fruit weight increased with increase of the organic fertilization doses up to 40 t/ha, reaching 1.05 kg for BRS Imperial (Figure 3) and 2.0 kg for the Pérola pineapple (Figure 4), both grown in the spacing of 1.20 m x 0.40 m x 0.40 m. The lower fruit weight obtained for the BRS Imperial cultivar was expected. In spite of its smaller fruits, BRS Imperial earned much higher fruit prices in the São Paulo market, probably due to its excellent taste and high total soluble solids (°Brix) content (Figure 5). In comparison to the Pérola cultivar, there is also the advantage of reducing production costs and increasing number of fruits harvested due to its resistance to the fusariosis disease.

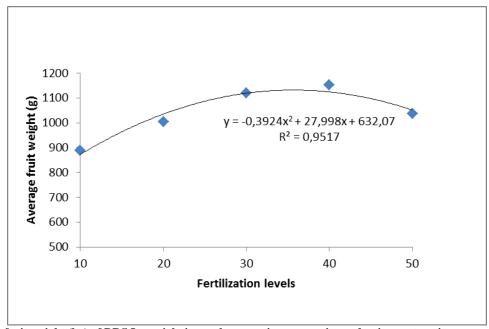


Figure 3. Average fruit weight (kg) of BRS Imperial pineapple grown in an organic production system, in response to five levels of organic fertilization (t/ha). Lençóis, Bahia, Brazil, 2015.

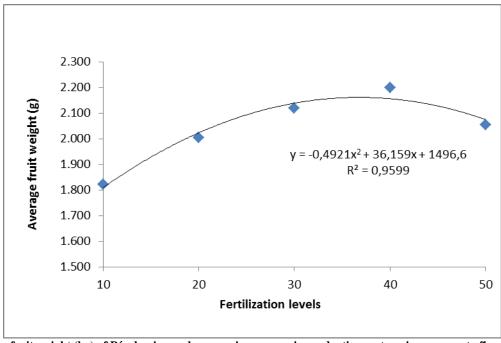


Figure 4. Average fruit weight (kg) of Pérola pineapple grown in an organic production system, in response to five levels of organic fertilization (t/ha). Lençóis, Bahia, Brazil, 2015.

The total soluble solids content (°Brix) of BRS Imperial pineapples (Figure 5) decreased with increasing level of fertilization, but was always superior to the minimum standard required for pineapple marketing in Brazil which is 12.0 °Brix.

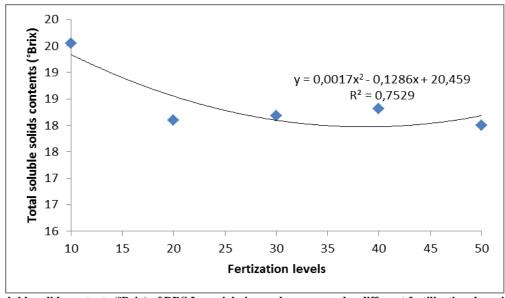


Figure 5. Total soluble solids contents (°Brix) of BRS Imperial pineapple grown under different fertilization doses in an organic production system. Lençóis, Bahia, Brazil, 2015.

There was no significant effect of fertilization levels on the vitamin C content, titratable acidity (TA) and <sup>°</sup>Brix:TA ratio of BRS Imperial fruits (Table 1). The results indicate a moderate acidity, lower than that of Smooth Cayenne pineapples, and high ratio which is higher than in Pérola fruits.

**Table 1** – Means of the physicochemical evaluations of Imperial pineapples grown under different fertilization

doses in an organic production system. Lençóis, Bahia, Brazil, 2015.

Doses	Vitamin C	Titratable Acidity	Ratio
(t/ha)	(mg/100g)	(% citric acid)	Kano
10	24.39a	0.55a	38.15a
20	27.33a	0.66a	28.35a
30	26.30a	0.54a	38.15a
40	25.64a	0.57a	35.06a
50	23.90a	0.36a	50.60a
Average	25.51 <sup>n.s</sup>	0.54 <sup>n.s.</sup>	38.06 <sup>n.s.</sup>

Values followed by same letters don't differ by the Tukey test at 5%

For Pérola pineapples, there was no significant dose effect on soluble solids or TA (Table 2) and the values were considered adequate for this cultivar.

Table 2 – Means of the physicochemical evaluations of Pérola pineapples grown under different fertilization

doses in an organic production system. Lençóis, Bahia, Brazil, 2015.

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Doses	Vitamin C	Soluble Solids	Titratable acidity
	(mg/100g)	(°Brix)	(% citric acid)
10	29.85	15.13	0.52
20	28.71	15.57	0.58
30	29.01	14.58	0.62
40	30.85	14.93	0.66
50	30.33	15.61	0.59
Average	29.75 <sup>n.s.</sup>	15.16 <sup>n.s.</sup>	0.59 <sup>n.s.</sup>

Values followed by same letters don't differ by the Tukey test at 5%

Planting density treatments did not significantly affect the fruit weight of BRS Imperial pineapples. Pérola, however, produced higher average fruit weights in the densities of 31,250 and 35,710 plants/ha (Table 3).

Table 3. Fruit weights (kg) of pineapple cvs. BRS Imperial and Pérola in response to different planting densities,

grown in an organic production system. Lencóis, Bahia, Brazil, 2015.

Planting densities	Pérola	BRS Imperial
(plants/ha)		
26,315	1.70 b	0.96 a
31,250	1.92 a	0.95 a
35,714	1.96 a	0.93 a
47,620	1.81 ab	0.95 a
51,282	1.84 ab	0.89 a
Mean	1.85	0.94
CV (%)	18.67	5.48

Values followed by same letters don't differ by the Tukey test at 5%

There was no influence of planting density on TA or soluble solids content of Pérola pineapple, with an acidity considered moderate for the pineapple crop (0.53%) and a °Brix of 15.0 considered high for this cultivar (Table 4).

Table 4. Means of the physicochemical evaluations of Pérola pineapples grown under different planting densities

in an organic production system. Lençóis, Bahia, Brazil, 2015.

Planting density (plants/ha)	Titratable acidity (% citric acid)	Soluble Solids (%)	Ratio	pН
26,315	0.56	15.1	28	3.69
31,250	0.50	15.6	31	3.69
35,714	0.53	14.8	29	3.74
47,620	0.52	15.2	30	3.74
51,282	0.56	14.7	27	3.71
Mean	0.53 <sup>ns</sup>	15.0 <sup>ns</sup>	29.0 ns	3.71 <sup>ns</sup>
CV (%)	8.94	3.98	7.91	1.44

There was no significant effect of planting density on the physicochemical characteristics of BRS Imperial pineapples (Table 5). Under the local cropping conditions the fruits of this cultivar had much higher average soluble solids content than did Pérola pineapple.

 Table 5 - Means of the physicochemical evaluations of BRS Imperial pineapples grown under different planting

densities in an organic production system. Lençóis, Bahia, Brazil, 2015.

Planting density	Titrabale	pН	Soluble solids	Ratio	Vitamin C
(plants/ha)	acidity (%	_	(°Brix)		(%)
	citric acid)				
26,315	0.49	4.11	20.00	42.70a	24.65ab
31,250	0.57	4.02	19.17	34.24ab	23.87ab
35,714	0.48	4.09	18.98	40.17ab	24.87a
47,620	0.48	4.23	19.89	44.54a	19.47c
51,282	0.62	4.07	19.87	32.18b	21.53bc
Means	0.53 <sup>ns</sup>	4.10 <sup>ns</sup>	19.58 <sup>ns</sup>	38.77	22.88
CV(%)	15.65	3.48	4.05	13.77	7.10

Regarding the management of soil cover plants done before planting the pineapple cultivars, the results indicate differences in the stock of nutrients in the biomass of these plants. Millet had the largest amount of phosphorus (P) and potassium (K) in the biomass, with more than 12 kg/ha P and 80 kg/ha K. Noteworthy is the potential for soil calcium extraction by the jack bean, above 140 kg/ha (Figure 6).

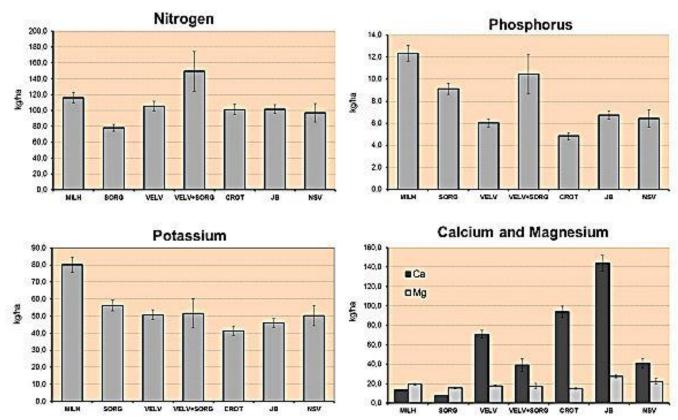


Figure 6. Macronutrient stocks in the biomass of different soil cover crops used as a pre-cultivation in organic production of pineapple. MILH, millet; SORG, sorghum; VELV, velvet bean; VELV + SORG, 1:1 blend of VELV & SORG; CROT: *Crotalaria juncea*; JB, Jack bean; NSV, spontaneous native vegetation. Horizontal bars indicate the standard error of the mean.

When analyzing the average weight of the pineapple fruits due to cover crops cultivation in pre-planting pineapple (Figure 7), results indicated higher production for Pérola pineapple cultivated in the spacing of  $1.20 \text{ m} \times 0.40 \text{ m} \times 0.40 \text{ m}$ , when combining velvet bean + sorghum which resulted in fruit weights above 2.3 kg. For all soil cover crop treatments the average fruit weight was more than 1.5 kg, which is the parameter for fruits of first quality in Brazil.

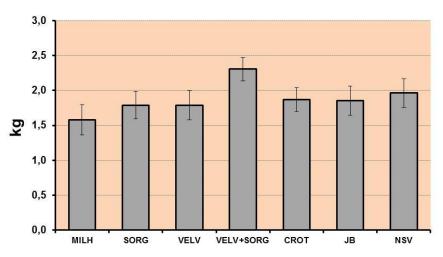


Figure 7. Average fruit weight of 'Pérola' pineapples in response to different soil cover plants used as pre-planting cultivation in an organic production system of pineapple. MILH, millet; SORG, sorghum; VELV, velvet bean; VELV + SORG, 1:1 blend of VELV & SORG, CROT, Crotalaria juncea; JB, Jack bean; NSV, spontaneous native vegetation.

Based on the above results, the main recommendations for the cultivation of BRS Imperial and Pérola pineapples in an organic production system in Lençóis, Bahia, Brazil are presented in Table 9.

**Table 9.** Recommendations for the cultivation of pineapple in an organic system for 'BRS Imperial' and 'Pérola' in the region of Lençóis, Bahia, Brazil.

Aspects of production and fruit	Cultivar				
characteristics	BRS Imperial	Pérola			
Soil preparation	Minimum Movement,	Minimum Movement,			
	planting of cover crops	planting of cover crops			
	(blend of grasses and	(blend of grasses and			
	legumes)	legumes)			
Planting density in double rows	35,714 to 47,617	35,714 to 51,282			
system (plants/ha and spacing)	(1.00 m x  0.40 m, 0.40 m;	(1.00m x 0.40m x)			
	1.00 m x  0.40 m x  0.30 m)	0.40m; 0.90m x 0.40m			
		x 0.30m)			
Fertilization at planting date and	Cattle manure, 300g +	Cattle manure, 300g +			
the first one on soil surface after	rock powder, 150g	rock powder, 150g			
planting (g/plant)					
Other post-planting fertilizations	Bokashi, 280 to 320g	Bokashi, 260 to 312g			
(5°, 7°, 10° months after					
planting; g/plant)*					
Average fruit weight (kg)	1.05	2.00			
Titratabale acidity (% citric	0.54	0.59			
acid)					
Total soluble solids ( <sup>o</sup> Brix)	19.00 to 20.00	15.00			
Ratio (soluble solids/acidity)	38 to 44	29			
Vitamin C content (mg/100g)	25	29.75			

<sup>\*</sup>Corresponding to 35 to 40 tons of fertilizer/ha for the 'BRS Imperial' and 25 to 30 tons of fertilizer/ha for 'Pérola'.

# Cultivation of Pineapple in Organic System – Recommendations Based on Studies in the Semiarid Region of the Chapada Diamantina, Bahia, Brazil

Tullio Raphael Pereira de Pádua, Aristóteles Pires de Matos, Ronielli Cardoso Reis, Eliseth Souza Viana, Raul Castro Cariello Rosa, Francisco Alisson da Silva Xavier, Embrapa Cassava & Fruits, Cruz das Almas, Bahia, Brazil

Pineapple cultivation in an organic production system is still a little explored activity and with little technical information available. In this cultivation system, the use of inputs is controlled by legislation and the farm must be certified by a control body. The use of conservation practices, products for biological control and minimal soil disturbance in crop management of this system results over time in an increase of biodiversity, reduction of losses by erosion and leaching of topsoil and an improvement of its physical and chemical characteristics.

A good planning for organic production is essential and the following aspects should be taken into account:

- Survey of potential markets for organic pineapple to be focused;
- Definition of the cultivar, planting system and planting time
- Choice of the planting area
- Physical and chemical analyses of the soil
- Definition of soil conservation and management practices in line with the principles of organic farming
- Maintenance and improvement of the physical, chemical and biological soil characteristics
- Identification of weeds in the area to be cultivated and of pests present in nearby pineapple crops
- Acquisition and selection of good quality planting material
- Monitoring and control of pests and natural enemies of those pests present in the area
- Number and dates of fertilizer applications after planting
- Planning the date for floral induction treatment and fruit harvest
- Adequate fruit packaging and transport to the markets

The use of varieties with resistance to the main diseases of the crop is always a great help when looking for an organic production system, as it eliminates the need of other control measures and reduces economic and environmental costs. However, it is not always possible to rely on resistant varieties depending on the market demand and there is always a chance to get adequate yields for cultivars susceptible to important pests. In the case of pineapple in Brazil, if a cultivar susceptible to *Fusarium* must be used, a strategy should be established for the control of this fungus, including practices such as monitoring and rouging, the use of *Fusarium* free plantlets obtained in nurseries from stem sections and an adequate planning of the floral induction to get flowering during a period with environmental conditions less favorable to the fungus.

With respect to soil management, conservation practices must be carried out focused on the improvement of its physical, chemical and biological properties. Among other practices that should be used are minimum tillage, cover crops and mostly flat areas. The area should initially be cultivated with a cocktail of soil quality enhancer plants including grasses and legumes (Table 1). Among the benefits of this practice are improvements in soil structure, nutrient cycling from the lower to the upper soil layers, an increased biodiversity and an increase in the organic matter and nutrient contents in the upper soil layer? The area should be prepared by a light weeding to facilitate sowing and broadcast fertilization of the soil quality enhancer plants. About 90 to 120 days after sowing, and before flowering, the plants should be mowed and the biomass kept on the soil surface.

**Table 1**. Composition of the cocktail to be used for soil preparation at pre-planting of pineapple in an organic production system in Northeast Brazil.

Species	% of the mixture	Seed amount (kg/ha)
Pork bean (Canavalia ensiformis)	25	30.0
Velvet bean (Mucuna aterrima)	25	22.5
Pearl millet (Pennisetum glaucum)	25	3.8
Sorghum (Sorghum bicolor)	25	7.5

The biomass should be allowed to dry over a 15-day period. Thereafter a light disking can be done in order to cut and incorporate the remaining crop residues or, if possible, pineapple may be planted directly into holes or grooves opened through the dried biomass.

The strict selection of the planting material is essential to prevent the entry of pests and for the success of the enterprise. Plantlets, slips or suckers at sizes of at least 30 cm must be obtained and selected from areas with no or low incidence of fusariosis. If possible, plantlets should be produced from pineapple stem sections (Figure 1), a system that assures clean planting material if correctly used.



Figure 1. Production of plantlets by stem sectioning of pineapple cultivars BRS Imperial and Pérola. (Photos by Pádua, T.R.P.)

After the initial growth and establishment of the pineapple crop, pests should be monitored monthly. Plants with symptoms of fusariosis should be removed and buried or burned to control the disease by reduction of the inoculum in the field. In the case of incidence of mealybugs, the transmitter of the pineapple wilt virus, soap syrup (Table 2) should be applied to the lower part of the plant. One week later should be observed if the treatment was efficient and, if necessary, another application should be done.

Table 2. A mixture of ingredients for the control of mealybugs. The mixture is to be diluted in water at a concentration of 0.5% to 3%.

Ingredients	Amount
Vegetal oil	3.7 liters
Ethanol	3.8 liters
Caustic soda	800 grams
Water	1.7 liters
Total	10 liters

During planting, each hole or groove should be fertilized with 350 grams of manure (nitrogen source) and 150 grams of powdered rock per plant (phosphorus source). After planting, topdressings with an organic fertilizer, split into 3 to 5 applications according to availability of material and skilled labor, should be placed at the base of the plant (Figure 2). In the case of three applications, the first topdressing should be done between months 2 and

3, the second between months 4 and 5, and the third between 9 and 10 months after planting. Potassium as potassium sulphate, which is in alignment with the present rules for organic production, should be applied along with organic compost or manure at dosages of 4.6, 6.2 and 7.0 g of potassium sulphate per plant.





Figure 2. Area of production of organic fertilizers type Bokashi and fertilization after planting directed to the lower part of the drip irrigated plants (Photos by Pádua, T.R.P.)

For weed control, cuttings should be done as often as necessary to avoid competition for nutrients, light and water between weeds and the pineapple plants, especially in the first six months of cultivation when the plant is more vulnerable and plant development may be affected and time span until harvest increased. Weed control in the organic system can be done via manual weeding with hoes or by using a power mower and through the use of plastic mulching.





Figure 3. Weed control by a coastal mower or by using plastic mulch. (Photos by Pádua, T.R.P.)

Floral induction in an organic pineapple production system can be done using calcium carbide as there is a permission for that by the certification body. Its application should occur in the late afternoon of less hot and preferably cloudy days. It is recommended to repeat the application 48 hours later, especially for the BRS Imperial pineapple. Calcium carbide should be diluted at a dosage of 400 g of calcium carbide in 100 liters of water, or if costal sprayers of 20 liters capacity are used, 50 to 60 g of calcium carbide should be placed into 12 liters of water leaving some space in the sprayer for the expansion of the acetylene gas released. 50 ml of the solution is applied directly into the central part of the leaf rosette of each plant. As fruit size is to a large extent related to plant size at forcing, this treatment should be done when 'D' leaves of plants present minimum fresh weights of 80 g and 60 g and minimum lengths of 100 cm for Pérola and 80 cm for BRS Imperial, respectively.

After the appearance of the first flowers the inflorescence should be covered using paper bags for protection against sunburn. This practice may also be a protection against *Fusarium* and the fruit borer when placed as soon as possible. Fruit harvesting should be done keeping about 2 cm of the flower stalk below the fruit

base in order to reduce the risk of rotting (Figure 4). Protection of the cut surface of the stalk by application of Bordeaux mixture is also recommended.



Figure 4. Aspects of a BRS Imperial pineapple field, its fruits and the presence of a short piece of the stalk at the fruit basis. (Photos by Pádua, T.R.P. (A e B) and Junghans, D.T. (C))

Fruits produced in an organic system usually have higher value in large markets, attending commercial niches that pay more depending on the quality of the product. To serve these markets, after harvest fruits should be packed into cardboard boxes that increase protection and reduce the risk of fruit losses during transport from the production area to the consumer center ensuring adequate fruit quality (Figure 5).



Figure 5. BRS Imperial and Pérola pineapples in cardboard boxes for long distance transport to consumer centers (Photos: Pádua, T.R.P. (A, B) and Matos, A.P.de (C))

# Use of Baits for Integrated Pest Management (IPM) of Ants, Pineapple Mealybugs, and Mealybug Wilt Disease of Pineapple in Espírito Santo, Brazil

M.P. Culik, D. dos S. Martins, and J.A. Ventura

Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural CRDR-CN, Linhares, Espírito Santo, Brazil

Keywords: Dysmicoccus brevipes (Hemiptera: Pseudococcidae), abamectin, Formicidae

#### Abstract

Research is being conducted in Espírito Santo, Brazil to evaluate the use of baits containing insecticides for control of ants associated with mealybugs infesting pineapple to reduce mealybug populations and mealybug wilt disease of pineapple. Initial results indicate that baits containing abamectin may be useful for control of ants and thus may help reduce mealybug populations and mealybug wilt incidence.

#### INTRODUCTION

As a vector of pineapple mealybug wilt-associated virus (PMWaV) and co-factor in development of mealybug wilt disease, the pineapple mealybug (*Dysmicoccus brevipes*) is a major pest of pineapple throughout the world including Brazil. Research in various regions indicates that use of baits containing insecticides to control of ants symbiotically associated with mealybug pests in pineapple and other crops may be effective in reducing ant and associated mealybug pest populations (Taniguchi et al., 2005; Daane et al., 2008). Therefore, we are currently conducting research to evaluate effects of baits for control of ants and associated mealybugs on pineapple and several other crops in Espírito Santo, Brazil, as part of continued development of integrated pest management (IPM) in this region

#### MATERIALS AND METHODS

#### **Preliminary tests**

Preliminary experiments to evaluate the effectiveness of baits containing the insecticides abamectin, boric acid, and spinosad for control of ants in pineapple fields and other crops (coffee and cocoa) have been established at the Instituto Capixaba de Pesquisa, Assistência Técnica e Extensão Rural (Incaper) Experiment Farm in Sooretama, and in Linhares, Espírito Santo.

The study sites consist of established fields. Twelve plants, separated from each other by at least 15 m were identified, and four plants in three blocks were randomly selected for each test treatment (Abamectin, Boric acid, Spinosad, and Untreated control). Granular baits for the tests were prepared with active ingredients (AI): abamectin (0.01%), boric acid (1%), or spinosad (0.015%). At the start of the test, for each bait treatment, bait (35 g) containing the AI was placed in a bait station (perimeter patrol) located at the base of each test plant (for boric acid treatments in pineapple and coffee an additional 35 g of the boric acid bait was added on the day following the initial application). Additional bait (35 g) was added for all treatments after 2 weeks.

To evaluate the effects of the baits on ants, ant activity was monitored at the start of the test and 2 and 4 weeks following bait application as follows: an open 50 ml plastic centrifuge tube with untreated bait (5 ml) was placed at the at the base of each test plant and after approximately one hour the tubes, with ants attracted to the bait inside, were closed and collected for transport to the laboratory for examination. The ants captured in the tubes were subsequently counted and preserved for identification and reference.

#### RESULTS AND DISCUSSION, CONCLUSIONS

Since this research has only recently begun results are limited at present. However based on our initial observations abamectin appeared to be most effective in reducing ant populations in the crops studied (Figure 1).

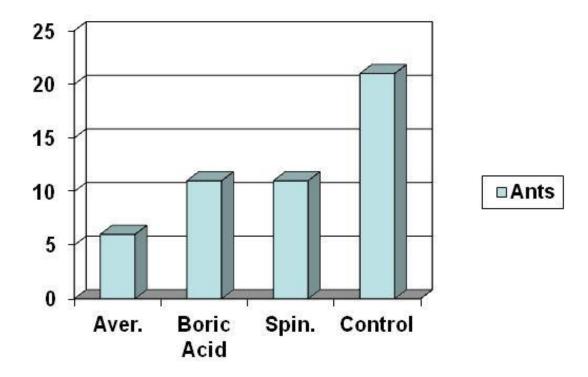


Figure 1. Effect of bait treatments containing abamectin (Aver.), boric acid, spinosad (Spin.) and untreated control on ants (mean number of ants/monitoring tube, 2 and 4 weeks after bait application) in cocoa, coffee, and pineapple crops, Espírito Santo, Brazil (Test 1, 2016).

#### **Field experiments**

Based on results of initial tests we plan to evaluate at least one active ingredient in baits in larger field trials for several crop seasons. We will compare pineapple plots with and without baits (10X10 m plots separated from each other by at least 10 m, with 3 replications) with bait stations placed on each side of treated plots, with ant activity recorded as described for the preliminary tests, and additional variables such as ant trailing activity, ant and mealybug infestation levels, and natural enemy activity also monitored.

#### **ACKNOWLEDGEMENTS**

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#### **Literature Cited**

Daane, K. M., Cooper, M. L., Sime, K. R., Nelson, E. H., Battany, M. C., Rust, M. K. 2008. Testing baits to control Argentine ants (Hymenoptera: Formicidae) in vineyards. Journal Economic Entomology 101: 699-709.

Taniguchi, G., Thompson, T., Sipes, B., 2005. Control of the big-headed ant, *Pheidole megacephala* (Hymenoptera: Formicidae) in pineapple cultivation using Amdro in bait stations. Sociobiology 45:1-7.

# Influence of Washing and Plastic Packaging on Post-harvest Quality of Ornamental Pineapple Stalks

Davi Silva Costa Júnior<sup>1</sup>, Everto Hilo de Souza<sup>2</sup>, Márcio Eduardo Canto Pereira<sup>2</sup>, Maria Angélica Pereira de Carvalho Costa<sup>1</sup> and Fernanda Vidigal Duarte Souza<sup>2\*</sup>

#### Introduction

Ornamental pineapple is a promising alternative in the segment of floriculture, as it can be marketed as cut flowers, potted plant, in landscaping, as a hedge or as ornamental minifruits (Souza et al., 2012). Ornamental pineapple floral stalks (crown, developing inflorescence or syncarp, peduncle and any bracts or leaves attached to it) are already produced and marketed in Brazil, mainly in the states of Ceará and Rio Grande do Norte (Brainer and Oliveira, 2007). For marketing they need to pass through standardization and post-harvest procedures, including the removal of flower debris, sprouts and leaves at the base of the peduncle and washing with high pressure water for the removal of scale-like trichomes, which are popularly and wrongly called wax.

The post-harvest management is one of the main problems faced by floriculture, where losses on the way between production and consumers can reach 50% (Teixeira, 2002). So the proper handling of the products is essential in order to keep quality, increase shelf life and reduce losses of inflorescences after cutting.

The floral stalks of ornamental pineapples are covered by trichomes with a powdery appearance, overshadowing much of its beauty, expressed mainly by its colors. However, the removal of these trichomes by pressure washing significantly increases the final cost of the product (Costa Júnior et al., 2011a). In the case of exporting companies, this procedure represents one of the main production costs. This process also tends to damage the most sensitive plant tissues, especially the bracts located under the syncarp, the crown and the bracts of the fruitlets, which in hybrids derived from *Ananas comosus* var . *bracteatus* are large and present pronounced colors (Costa Júnior et al., 2011b).

According to Chitarra and Chitarra (2005) methods to reduce postharvest losses include the application of modified atmosphere, which can be generated by the use of plastic films or edible coatings that induce changes in the atmosphere around the product by increasing the concentration of  $CO_2$  and decreasing that of  $O_2$  due to respiration of the product. However, the selective permeability of the film must be suitable for the input of  $O_2$  and the output of  $CO_2$  to avoid anaerobic conditions.

The objective of this study was to evaluate the post-harvest life of flower stalks of an ornamental pineapple hybrid developed by Embrapa Cassava and Fruits under different conditions of packaging.

#### Methodology

Floral stalks of the ornamental pineapple hybrid PL04 were used. This hybrid was originated from a cross between *A. comosus* var. *erectifolius* and *A. comosus* var. *bracteatus*. Stalks having a length of 35 cm were cut at their base after closure of the last flower, but not older than three days after that time. Buds at the base of the syncarp as well as leaves present at the base of the peduncle were removed. Trichomes on the stalks were removed by an electric high pressure washer to achieve commercial standard for the export market. Water jet length was kept at 20 cm distance from the stalk surface. Thereafter the stalks were dried in the shade at room temperature and randomly divided into groups for treatment application.

In order to determine the effect of washing and low density polyethylene (LDPE) packaging on postharvest quality of the floral stalks the following treatments were studied:

- T1) Stalk without washing (control)
- T2) Washed stalk
- T3) Stalk not washed + full packaging with 4 uM LDPE
- T4) Stalk not washed + full packaging with 8 um LDPE
- T5) Stalk not washed + partial packaging with 8 μm LDPE

<sup>&</sup>lt;sup>1</sup> Universidade Federal do Recôncavo da Bahia, 44380-000, Cruz das Almas, Bahia, Brazil

<sup>&</sup>lt;sup>2</sup>Embrapa Cassava & Fruits, 44380-000 Cruz das Almas, Bahia, Brazil

<sup>\*</sup> Corresponding author: fernanda.souza@embrapa.br

- T6) Washed stalk + full packaging with 4µm LDPE
- T7) Washed stalk + full packaging with 8 μm LDPE
- T8) Washed stalk + partial packaging with 8 µm LDPE

The LDPE plastic bags were 10 cm wide and 15 cm long and 4  $\mu$ m or 8  $\mu$ m thick. In the full packaging treatments two bags were used, one placed from the top of the crown downward and the other from the peduncle base upward warranting a full covering of crown, syncarp and peducncle. In the partial packaging treatments one bag was used covering only crown and syncarp (Figure 1). After application of the treatments the stalks were arranged in floral sponges and stored at room temperature.



Figure 1. Ornamental pineapple stalks fully or partially packed with 8 µm low density polyethylene (LDPE).

The overall appearance of the floral stalks and the specific appearance of each part (crown, syncarp, peduncle) were evaluated individually every two days during the storing period of 24 days, using the following scale: 4 = Excellent: vivid color, shiny and absence of dryness; 3 = Very good: lightly faded color or lightly yellowed and some early drying at the extremities; 2 = Good: more faded and yellowed color and drying affecting about 1/3 of the surface; 1 = Bad: partially degraded colors and drying of about 2/3 of the surface; and 0 = Very bad: Very degraded color (colorless or brown) and completely dry.

The average grade equal to 3 (very good) for appearance of the different parts of the floral stalk was defined as the limit of shelf life for marketing. The experimental design was a completely randomized one using eight treatments with eight replications. Data were submitted to analysis of variance and means were compared by the Scott-Knott test at 5% probability using the SAS statistical software (SAS Institute, 2010).

#### **Results and Discussion**

There was significant difference between treatments for the four variables studied. The best overall result was obtained by using a full packaging of 8 µm thickness, with or without washing (treatments 4 and 7), which extended stalk shelf life by four days (Table 1). The unwashed and not packed (control) stalks were evaluated with grade 3 (very good) for a shelf life of up to 10 days, while the washed ones (T2) were in good shape for only six days, showing a negative effect of washing accelerating color fading and overall drying.

The 8  $\mu$ m LDPE (T4, T7 – see bold characters) significantly increased shelf life by 8 days compared to the 4  $\mu$ m LDPE (T3, T6) treatment. Packaging, with or without washing, improved overall general performance of all floral parts. These results show that although washing is a procedure that may cause tissue damage, the use of packaging protecting the entire stalk, reducing water loss due to the microclimate established inside the bags. Crowns achieved longer shelf life when compared to the appearance of peduncle and syncarp. Despite some additional costs which may affect the final price of the product, a shelf life extension by a few days as observed in this study may be a valuable advantage making the investment worth for exporters.

**Table 1.** Evolution of post-harvest shelf life of floral stalks of the ornamental pineapple hybrid PL04 (*Ananas comosus* var. *erectifolius* X *A. comosus* var. *bracteatus*) in response to washing and LDPE packaging treatments

Treat <sup>†</sup>	us vai.				n period	. <i>bracted</i> (davs)	11113) III I	response	to was	ining and	LDIE	packagi	ng treat	Sig <sup>‡</sup>
	0	2	4	6	8	10	12	14	16	18	20	22	24	8
		Flora	al stalk c	overall a	ppearanc	e								
T1	4	4	3	3	3	3	2	2	2	1	1	1	0	C
T2	4	4	3	3	2	2	2	1	1	1	1	0	0	C
T3	4	4	3	3	2	2	2	2	2	2	1	1	0	D
T4	4	4	3	3	3	3	3	3	2	2	1	1	0	$\mathbf{A}$
T5	4	4	3	3	3	3	3	2	2	1	1	1	1	В
T6	4	4	3	3	2	2	2	2	2	2	1	1	0	D
T7	4	4	3	3	3	3	3	3	2	2	1	1	0	$\mathbf{A}$
T8	4	4	3	3	3	3	3	2	2	1	1	1	0	В
CV %														5.32
		Pedu	incle app	oearance										
T1	4	4	4	3	3	3	2	2	1	1	1	0	0	C
T2	4	4	4	3	3	3	2	1	1	1	1	0	0	C
T3	4	4	4	3	3	3	2	2	2	1	1	0	0	C
T4	4	4	4	4	3	3	3	3	2	2	1	0	0	$\mathbf{A}$
T5	4	4	4	4	3	3	3	2	2	1	1	0	0	В
T6	4	4	4	3	3	3	2	2	2	1	1	0	0	C
T7	4	4	4	4	3	3	3	3	2	2	1	0	0	$\mathbf{A}$
T8	4	4	4	4	3	3	2	2	2	1	1	0	0	C
CV %														8.32
		Sync	carp appe	earance										
T1	4	4	4	3	3	2	2	2	1	1	1	1	0	C
T2	4	4	3	3	2	2	1	1	1	0	0	0	0	C
T3	4	4	4	3	3	2	2	2	2	1	1	1	0	C
T4	4	4	4	4	3	3	3	2	2	2	1	1	0	$\mathbf{A}$
T5	4	4	4	4	3	3	3	3	2	2	1	1	0	$\mathbf{A}$
T6	4	4	4	3	3	2	2	2	1	1	1	1	0	C
T7	4	4	4	3	3	3	3	2	2	1	1	1	0	$\mathbf{A}$
T8	4	4	4	4	3	3	2	2	2	1	1	1	0	В
CV %														8.21
		Crov	vn appea	arance										
T1	4	4	4	4	4	3	3	3	3	3	2	2	1	C
T2	4	4	4	4	3	3	3	3	3	2	1	1	1	D
T3	4	4	4	4	3	3	3	3	3	2	2	2	1	D
T4	4	4	4	4	4	4	4	4	4	4	4	3	2	$\mathbf{A}$
T5	4	4	4	4	4	4	4	4	4	4	3	2	2	В
T6	4	4	4	4	3	3	3	3	3	2	2	1	1	D
T7	4	4	4	4	4	4	4	4	4	4	4	3	3	$\mathbf{A}$
T8	4	4	4	4	4	4	4	4	4	4	3	3	2	$\mathbf{A}$
CV %														6.21

<sup>†</sup>Treatments are T1, Control; T2, washed; T3, not washed + full 4 μm LDPE packaging; T4, not washed + full 8 μm LDPE packaging; T5, not washed + partial 8 μm LDPE packaging; T6, washed + full 4μm LDPE packaging; T7, washed + full 8 μm LDPE packaging; T8, washed + partial 8 μm LDPE packaging. Scale used: 4 = Excellent; 3 = Very good; 2 = Good; 1 = Bad; 0 = Very bad.

#### References

Brainer, M. S. C. P.; Oliveira, A. A. P. 2007. Floricultura: perfil da atividade no Nordeste Brasileiro. Fortaleza: Banco do Nordeste. 351p. (Documentos do ETENE, n.17).

<sup>&</sup>lt;sup>‡</sup>Treatments followed by the same letters within a category are not significantly different by Scott-Knott test at 5 %.

- Cavalcante, R. A.; Mosca, J. L.; Paiva, W. O.; Maciel, V. T.; Almeida, J. B. S. A.; Guimarães, A. A. 2007. Conservação pós-colheita de Sorvetão (*Zingiber spectabile* Griff.) utilizando filme plástico em diferentes pontos de colheita. Revista Brasileira de Horticultura Ornamental, v.12, p.117-121.
- Chitarra, M. L. F.; Chitarra, A. B. 2005. Pós-colheita de frutos e hortaliças fisiologia e manuseio. Lavras: UFLA. 785p.
- Costa Júnior, D. S.; Pereira, M. E. C.; Souza, F. V. D.; Carvalho, H. L. 2011a. Remoção da cera natural de hastes florais de híbridos de abacaxi ornamental. In: Simpósio de Pós-Colheita de Frutas, Hortaliças E Flores, 3., 2011, Nova Friburgo. Anais..., Nova Friburgo: Embrapa Agroindústria de alimentos, 2011, p. 79-82.
- Costa Júnior, D. S.; Pereira, M. E. C.; Souza, F. V. D. 2011b. Longevidade pós-colheita de hastes florais de híbridos de abacaxi ornamental tratadas com soluções de condicionamento. In: SIMPÓSIO DE PÓS-Colheita De Frutas, 25 Hortaliças E Flores, 3., 2011, Nova Friburgo. Anais..., Nova Friburgo: Embrapa Agroindústria de alimentos, 2011 p. 83-86.
- SAS Institute. 2010. SAS user's guide: statistic: version 9.2. Cary: SAS Institute.
- Souza, E. H.; Souza, F. V. D.; Costa, M. A. P. C.; Costa Júnior, D. S.; Santos-Serejo, J. A.; Amorim, E. P.; Ledo, C. A. S. 2012. Genetic variation of the *Ananas* genus with ornamental potential. Genetic Resources and Crop Evolution, v. 59, p. 1357-1376.
- Teixeira, M. C. F. 2002. Curso prático de pós-colheita para flores tropicais. In: Antunes, M. G. Floricultura em Pernambuco. Recife: SEBRAE, p.11-15.

# **News from Costa Rica**

# **Burndown of Pineapple Plants in Costa Rica**

Jhonny Vásquez Jiménez . San Carlos, Costa Rica. E-mail: jvasquez@proagrocr.com

Herbicide burndown of pineapple plantation fields is an agricultural practice for eradicating plants that have completed their growing cycle. It is done in order to start preparing the soil for planting a new crop. It is considered a good result when a burndown of 80% of the plant material is achieved (Garita, 2014). Currently, the practice is performed relatively successfully with the use of 6 to 9 L ha<sup>-1</sup> (commercial product) of paraquat herbicide (recommendation of some paraquat 20 SL label's ). Despite the success achieved by Costa Rica companies employing this practice, they have been challenged on the use of paraquat for more than 5 years. Therefore, there is a need to replace this herbicide by others that are not questioned by buyers in fruit markets.

#### **Burndown substitute herbicide characteristics**

The desirable characteristics of a burndown herbicide are:

- Desiccate pineapple plants as quickly as possible, depending on weather conditions (the standard of paraquat is 30 days after application), Garcia and Rodriguez (2009).
- The foliage and the mother stem must be killed because buds growing on the stem of pineapple plants will result in undesirable volunteer suckers in the new planting.
- The kill must be rapid enough so that it does not support the reproduction of the stable fly (*Stomoxys calcitrans*), since pineapple crop residues are suitable for stable fly reproduction (Pitta, 2011).
- The quantity of active ingredient per hectare needs to be less than that for paraquat, i.e., less than 1.2 1.8 kg ha<sup>-1</sup> a.i.

The 30 days criteria for plant desiccation may be undesirable for some producers with short inter-cycles, but would not be a problem for companies that follow best practices of soil conservation. For such companies, it could be considered a technical advantage because the soil would be covered with stubble while the residue of the last crop was drying.

Since 2012 we have been evaluating the biological efficacy of pineapple burndown by herbicides containing imazamox as the active ingredient. The first tests were carried out on plantations of northern Costa Rica, where several active ingredients were tested, all at the highest dose according to the label. The objective was to assess whether any of the active ingredients tested had the characteristics of a burndown herbicide for use on pineapple. We found that imazamox had interesting behavior, but it was necessary to work in many other doses.

In the following years specific tests were run to determine the dose of active ingredient able to get the desired burndown of the pineapple crop in scenarios consistent with the cultivation practices in Costa Rica. Our results to date with imazamox are summarized below.

We obtained 80% burndown between 45 and 60 days after application of imazamox to pineapple depending on the area (North Zone 45 days and Atlantic Zone 60 days; Figure 1A and 1B).

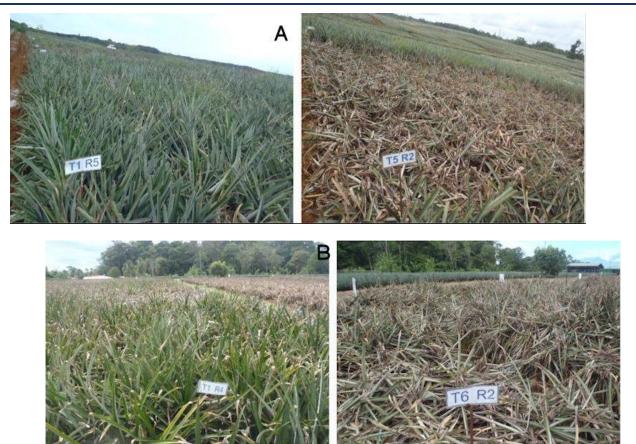


Figure 1A & B. Effects of imazamox herbicide on a pineapple field designated for burndown. Photo A (upper) shows the test in the Atlantic zone at 60 days after application. Photo B (lower) shows the test in the North zone at 45 days after application. The photoes on the left are of untreated fields while the field plots on the right were sprayed with a solution containing  $0.35~{\rm g\cdot L^{-1}}$  imazamox.

The biological efficacy of the active ingredient is achieved at a minimum concentration of  $0.35~g~L^{-1}$  a.i. when the average green biomass per plant is 1.5~kg (Figure 2). However, when the amount of green biomass present is 3.5~kg per plant or higher, a greater spray volume of  $4000~L~ha^{-1}$  must be used (Figure 3). All imazamox sprays included  $20~kg~ha^{-1}$  urea and  $0.3\%~DASH^{\$}$  adjuvant.



Figure 2. Effect of low biomass (less than 1.5 kg of green matter per mother plant and suckers) on burndown. Left (T1, R3) is the control, center shows the condition of T1 prior to spraying. At right is the burndown at 45 days after the application of 500 L ha<sup>-1</sup> L of spray containing 0.35 g L-1 a.i. of imazamox. Only 0.175 kg ·ha-1 a.i. of the herbicide was applied.



Figure 3. Effect of imazamox on burndown when the average green plant biomass was >3.5 kg (mother plant and suckers). At left is the untreated control, at center a mother plant and suckers and at right is the burndown achieved 60 days after applying the equivalent of 4000 L ha-1 of a spray containing 0.35 g L-1 a.i. of imazamox. (equivalent of 1.4 kg ha-1 a.i.).

At 15 to 30 days after application of imazamox there was progressive phytotoxic injury in the white basal tissue (leaf rosette), the leaves fold at this level and slowly begin to lose turgor until completely dried. This biocidal effect takes approximately 45 days in the North zone and about 60 days in the Atlantic zone, with the difference being due primarily to differences in the amount of biomass present in the two regions.



Figure 4. Progressive drying of pineapple plants after application of imazamox at, from left, 15, 30 and 45 days.

When evaluations of burndown were carried out over time (Figure 5), at 90 days after application, we found areas of bare soil present in the treatments with the recommended doses.



Figure 5. Advance of burndown in a pineapple field treated with 1.4 kg ha-1 a.i. imazamox in 4000 L of water. The photos above from left to right show the burndown at 25, 35, and 45 days after application (DAA) and similarly below at 60, 75 and 90 DAA.

When the recommended doses (concentration of active ingredient and volume of water) were used there was no evidence of regrowth even 90 days after application (Figure 6). However, if less water was used in areas with high biomass present, some buds were observed on the stems of mother plants.



Figure 6. Photos of plants at 90 days after spraying imazamox in a field where the average green biomass per plant was 3.5 kg. The left photo shows the absence of suckers at 90 days after spraying 1.4 kg h-1 a.i. of imazamox in 4000 L of water. The right photo is a plant from a plot in the same field sprayed with only 0.175 kg ha-1 a.i. of imazamox in 500 L of water (Note succulent leaves and presence of a developing bud (circled)).

Preliminary observations indicated there was a lower incidence of stable fly (*Stomoxys calcitrans*) in the burndown with imazamox, which was associated with fewer infection foci (Figure 7). When paraquat is used to burndown plant residue it creates "macerating" or watery tissues that attract flies. However, with imazamox we found only macerated stems of suckers present after imazamox was applied, but no fly larvae were found in the stem of the parent plant or in leaf tissue.



Figure 7. Changes in pineapple plants from 0 to 90 days after spraying 1.4 kg ha-1 imazamox in 4000 L of water. The average green plant biomass was 3.5 kg and at and after 45 days after application the leaf rosette and upper portions of the stem had begun to decay and produce odors that made the plants susceptible to invasion by the stable fly (Stomoxys calcitrans) (see arrows); however, no evidence was found of insect colonization.

Based on these investigations the process of registering Imazamox began in February 2016 (as an extension of uses "in the label" to include post-harvest burndown of pineapple) in Costa Rica by the "Servicio Fitosanitario del Estado". Resolution document: DOR-UBSI-PEB-026-2016.

Finally, it is appropriate to comment that some producers with particular conditions (specific targets and at specific times in the annual planting) have chosen methods of nonchemical break down, such as the use of effective microorganisms (EM's) to promote plant decomposition. They have achieved significant success in both the control of stable fly and the incorporation of stubble on the ground.

#### Acknowledgement

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#### Literature cited

García, A; Rodríguez, M. 2009. Reduciendo el Escurrimiento de Plaguicidas hacia el mar Caribe: Manual de buenas prácticas agrícolas para la producción de piña en Costa Rica. (en línea). Venecia, Costa Rica. Consultado 03 mar. 2015. Disponible en: <a href="http://cep.unep.org/repcar/proyectos-demostrativos/costa-rica-1/publicaciones-banacol/Manual%20BPA%20Banacol.pdf">http://cep.unep.org/repcar/proyectos-demostrativos/costa-rica-1/publicaciones-banacol/Manual%20BPA%20Banacol.pdf</a>

Garita, RA. 2014. La Piña. Cartago, Costa Rica: Editorial Tecnológica de Costa Rica. 568p.

PITTA (Programa de Investigación y Transferencia de Tecnología Agropecuaria, CR). 2011. Guía de diagnóstico de la mosca del estable Stomoxys calcitrans y otros dipeteros asociados a rastrojos. Costa Rica. Fitacori. 28 p.

# **News from Cuba**

# Hardening of 'MD-2' Micropropagated Pineapple Plants by Drought to Improve the Acclimatization-field Transition

René C. Rodríguez-Escriba<sup>1\*</sup>, Gustavo Y. Lorente<sup>1</sup>, Ibraín D. Rodríguez-Cartaya<sup>2</sup>, Dariel López<sup>1</sup>, Roberto E. Izquierdo<sup>1</sup>, Lucía S. Pérez-Borroto<sup>1</sup>, Pedro Guerrero Posada<sup>3</sup>, Carlos E. Aragón<sup>1</sup>, Camilo Bonet<sup>3</sup>, Yolanda Garza-García<sup>4</sup>, Romelio Rodríguez<sup>1</sup>, Florencio E. Podestá<sup>5</sup> and Justo L. González-Olmedo<sup>1</sup>.

<sup>1</sup>Laboratorio de Agrobiología. Centro de Bioplantas. Universidad de Ciego de Ávila. Ciego de Ávila, Cuba. <sup>2</sup>Facultad de Biología de la Universidad de La Habana. La Habana, Cuba. <sup>3</sup>Instituto de investigaciones de Ingeniería Agrícola. La Habana, Cuba. <sup>4</sup>Departamento de Biotecnología, Universidad Autónoma de Coahuila. Coahuila, México. <sup>5</sup>Centro de Estudios Fotosintéticos y Bioquímicos, Universidad Nacional de Rosario. Rosario, Argentina.

\* To whom correspondence should be addressed: e-mail: renecarlos@bioplantas.cu

#### **Abstract**

Current technology of pineapple micropropagation in Cuba has problems with the plant transition from acclimatization stage to the field. These problems are associated with the abrupt environmental changes between the two growth conditions. This work describes the study of crassulacean acid metabolism (CAM) response of 'MD-2' micropropagated pineapple plants under water stress. Plants acclimated to high light intensity were exposed to drought (droughted) for 30 days, after which the plants were fully watered from day 30 to day 45. Control plants were well watered for the entire 45 day period. Droughted plants had a higher succulence index (SI), water use efficiency (WUE), CO<sub>2</sub> uptake ratio and superoxide dismutase (SOD; EC 1.15.1.1) activity and lower chlorophyll (Chl.) than control plants. Both plant groups showed a typical CAM performance, but this was stronger in droughted plants. However, at 15 days after droughted plants were fully watered Chl. increased and SI, WUE, CO<sub>2</sub> uptake ratio and SOD activity decreased. The quick recovery of the droughted plants shows a high metabolic plasticity to water stress and a better response to the acclimatization-field transition after 30 days of drought as long as they were watered during or after their planting in the field.

### INTRODUCTION

After bananas and mangoes the pineapple is the third most important tropical fruit produced in the world {FAOSTAT, 2013 #468} and the most important CAM plant {Borland, 2009 #244}. Traditionally the pineapple is propagated using crowns, slips, shoots or suckers, but these planting materials have limitations such as less uniformity, transmission of diseases, and inadequacy for commercial production {Bartholomew, 2009 #765}. *In vitro* propagation is a crucial technique for the rapid production of disease-free pineapple plantlets {Escalona, 2003 #1567}. However, *in vitro*-cultured plants are very sensitive to abrupt environmental changes, mainly when they are removed from *in vitro*-culture containers and placed under *ex vitro* conditions, and later from vessels to field plantation. Therefore, it is often necessary to adapt the plantlets to the harsher and uncontrolled *ex vitro* environment of a greenhouse or the field. This transitional phase of plant development is often called acclimatization or hardening {Rodriguez, 2008 #1564}. Plant hardening can be achieved through management of container size, substrates, and fertilizers, in addition to the control of light intensity, temperature and irrigation. Management of these issues can elicit metabolic responses after vegetative developments that are indispensable for their successful field transition.

Pineapple is a CAM (Crassulacean Acid Metabolism) plant {Bartholomew, 2003 #104}. CAM may operate in different modes which depends mainly of their CO<sub>2</sub> exchange pattern and nocturnal acid accumulation, and they are classified according to the response of both variables {Herrera, 2009 #88;Lüttge, 2004 #216}. Macro-propagated pineapple plants in the field or in their natural habitat are classified as "Strong constitutive CAM plants" {Bartholomew, 2003 #104;Nelson, 2008 #201;Lüttge, 2010 #113;Prigge, 2014 #135}. However,

micropropagated pineapple plants have a high metabolic plasticity, responding as C3 plants in *in vitro*-culture conditions and in the first weeks of *ex vitro* acclimatization. Their response can be switched to CAM through stress signaling.

Pineapple plantlets grown under *in vitro* conditions show CAM activity when grown in an alternating warm day-cool night temperature regime {Nievola, 2005 #98} and also by nitric oxide and water deficit applications {Freschi, 2009 #442}. Pineapple plantlets also can be conditioned to function as C3 or CAM plantlets during the first weeks of *ex vitro* growth by low relative humidity, high temperatures and high light intensity {Aragón, 2012 #60;Aragón, 2013 #137}. For these reasons we conjecture that micropropagated pineapple plants as presently grown *in vitro* do not have the magnitude of CAM expression or the defensive mechanisms against oxidative stress required to survive their transition to the field. Two key aspects of acclimatization-technology are easy and practical handling and the low cost production. Rodríguez-Escriba *et al.* (2015) improved the plant acclimatization-field transition process by exposing plantlets to direct sunlight for 30 days. Pineapple plant transition to the field could also be improved by controlled drought, which could induce the plants defensive mechanisms against water stress. The aim of this work was to evaluate the CAM response of micropropagated pineapple plants under water deficit for 30 days, and their subsequent recovery after fifteen days with irrigation.

#### MATERIAL AND METHODS

#### Plant material and growth conditions

Plant material (*Ananas comosus* (L.) Merr. 'MD-2') was micropropagated and acclimatized as described by Rodriguez-Escriba et al. (2015). After five months in acclimatization stage plants with 11-12 leaves, 12-13 roots and a FW of 34-36 g were selected in September, 2014 and acclimated to direct sunlight (Figure 1 A-D),  $700\pm60~\mu\text{mol m}^{-2}\text{s}^{-1}$  at 12:00 h, for 30 days {Rodríguez-Escriba, 2015 #1568} until November. Relative humidity and temperature were  $56\pm2~\%$  and  $36\pm3~\%$ C at midday and  $79\pm4~\%$  and  $26\pm3~\%$ C at midnight. Irrigation treatments, which were done with a watering can, are described in Table 1.



Figure 1. Micropropagated pineapple plants in aclimatization stage inside the greenhouse (A and B) and acclimated to high light intensity under direct sunlight (C and D).

**Table 1.** Growth conditions of micropropagated pineapple plants.

Treatments	Until 0 days	Between 0-30 days	After 30 days
Control plants	Well-watered (1)	Well-watered	Fully-watered (3)
Droughted plants	Well-watered	Non-watered (2)	Fully-watered

<sup>(1) 120</sup> mL per plants each two days at 9:00 h; (2) Drought; (3) 480 mL per plants every day at 9:00 h

#### **Morphological measurements**

Thirty plants were taken for the measurement of leaf number and D-leaf length and width. The D-leaf was the tallest leaf on plants (Figure 2) {Bartholomew, 2003 #104}. Fresh and dry weight (FW and DW respectively) were determined on discs cut from the middle part of four D-leaves taken at 15:00 h. The dry weight was obtained after drying the discs to a constant weight at 70 °C. D-leaf water content was calculated as the difference between fresh and dry weight. Results were expressed as g cm<sup>-2</sup>.

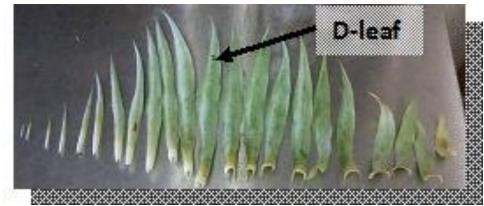


Figure 2. Leaf distribution according to their physiological age with the younger leaves on the left.

### **Gas Exchange measurement**

Gas exchange was done using a Portable Photosynthesis System (PP Systems CIRAS-2). Measurements were done on the middle part of four D-leaves which were enclosed in a rice leaf chamber (1.7 cm<sup>2</sup>). Data were collected from measures done each three hours during a whole day according to Rodríguez-Escriba et al. (2015).  $CO_2$  uptake ratio was calculated as the ratio net  $CO_2$  uptake between 00:00 h and 06:00 h: net  $CO_2$  uptake during the whole day (24 h). Water-use efficiency (WUE) was calculated as the ratio net  $CO_2$  uptake day<sup>-1</sup>: transpiration (T) day<sup>-1</sup>, µmol mmol<sup>-1</sup>.

#### Organic acid levels

Samples (1 g) from the middle of the D-leaf were stored under liquid nitrogen until determinations were made. Samples of four plants were added into a test tube containing 50% ethanol and were boiled at 80 °C for 20 min. The samples were titrated using NaOH [20 mM] and phenolphthalein as indicator. Organic acid levels (OA levels) were expressed as  $\mu eq. g^{-1} FW$ .

### Chlorophyll analyses and succulence index

Discs were cut on the middle part of three D-leaves taken at 15:00 h. Samples were stored under liquid nitrogen until determinations were made. Extraction and quantification was done as described by Rodríguez-Escriba et al. (2015) and results were expressed as µg cm<sup>-2</sup>. Succulence index (SI, water mass: Chl.<sup>-1</sup>, g mg<sup>-1</sup>) was calculated according to Kluge and Ting (1978).

### Extraction and quantification of protein and superoxide dismutase assay

Four D-leaves were collected at 15:00 h and were stored under liquid nitrogen until each determination was made. Protein extraction and quantification and superoxide dismutase (SOD) assay were done according to Rodríguez-Escriba et al. (2015).

#### Data analysis

Data were analyzed using the statistical software package Statistica v. 8.0 (Statsoft Inc., Tulsa, OK, USA). Before carrying out statistical tests normality of the data was checked by means of the Kolmogorov–Smirnoff statistic (P >0.05) and variances homogeneity by means of Levene's test (P >0.05). Means was compared by ANOVA factorial and Tukey's test ( $p \le 0.05$ ).

#### RESULTS AND DISCUSSION

#### Morphological response

After 30 days the droughted plants had fewer leaves, shorter D-leaf length and a slightly greater width than control plants (Table 2). The D-leaf water content of droughted plants was 2.2 times lower than that of control plants. However, 15 days after droughted plants were irrigated (45 days), D-leaf length had increased and water content was comparable to that of control plants. There were small increases in leaf number and D-leaf

width. The slightly greater D-leaf width could be the characteristic morphological response of micropropagated pineapple plants to stress signalling (Table 2). Micropropagated pineapple plants without previous treatments to improve field transition had an increase of the D-leaf width from 2.5 to 3.5 cm after 90 days of planting in the field {Rodríguez, 2013 #121}.

**Table 2.** Morphological response of micropropagated pineapple plants to water deficit.

Treatments	Leaf number		D-leaf length (cm)		D-leaf width (cm)		Water content of D- leaf discs (g cm <sup>-2</sup> )	
	30 days	45 days	30 days	45 days	30 days	45 days	30 days	45 days
Control plants	16.03 <sup>ab</sup>	16.52 <sup>a</sup>	24.69 <sup>b</sup>	26.62 <sup>a</sup>	3.35°	3.45 <sup>bc</sup>	1.44 <sup>b</sup>	1.64 <sup>a</sup>
Droughted plants	14.86 <sup>c</sup>	15.52 <sup>bc</sup>	$23.02^{c}$	$24.48^{b}$	3.53 <sup>ab</sup>	3.56 <sup>a</sup>	$0.65^{c}$	1.62 <sup>a</sup>
SE	0.92*		0.90*		0.0	)2*	0.01*	

Different letters and the symbol \* indicates significant differences among the four means within a category. Data were compared using ANOVA factorial followed by Tukey's test ( $p \le 0.05$ ) n=30 plants for leaf number and D-leaf length and width, n=4 plants for water content measurements.

## Gas exchange response

After 30 days of drought, D-leaf  $CO_2$  exchange and transpiration rates of the droughted plants between 00:00 and 06:00 h had values 3.4 and 5 times, respectively, lower than values for control plants (Table 3). However, leaves of droughted plants had the highest values of water-use efficiency (WUE) and  $CO_2$  uptake ratio. The  $CO_2$  uptake ratio between 00:00 and 06:00 h for droughted plants at 30 days was 99% of  $CO_2$  uptake during the whole day.

**Table 3.** Gas exchange response of micropropagated pineapple plants to water deficit between 00:00 and 06:00 h.

Treatments	CO <sub>2</sub> exchange rate (A) (µmol CO <sub>2</sub> m <sup>-2</sup> s <sup>-1</sup> )		Transpiration rate (T) (mmol H <sub>2</sub> O m <sup>-2</sup> s <sup>-1</sup> )		WUE (A/T) (µmol mmol <sup>-1</sup> )		CO <sub>2</sub> uptake ratio respect to the whole day	
	30 days	45 days	30 days	45 days	30 days	45 days	30 days	45 days
Control plants	6.92 <sup>b</sup>	7.31 <sup>ab</sup>	1.50°	1.91 <sup>a</sup>	4.73 <sup>b</sup>	3.85°	$0.80^{b}$	0.77°
Droughted plants	$2.03^{c}$	$7.98^{a}$	$0.31^{d}$	1.71 <sup>b</sup>	$6.60^{a}$	4.74 <sup>b</sup>	$0.99^{a}$	$0.79^{bc}$
SE	0.03*		0.01*		0.08*		0.01*	

Different letters and the symbol \* indicates significant differences among the four means within a category. Data were compared using ANOVA factorial followed by Tukey's test ( $p \le 0.05$ ). Analyses were done to n=4 plants. Data are means of measures done each three hours between 0:00 and 6:00 h except to  $CO_2$  uptake ratio which is the result of the mean of these hours respect to the mean of the whole day.

When fully watered for 15 days, droughted plants recovered rapidly and had a normal CO<sub>2</sub> exchange rate while the transpiration rate was still significantly less than for leaves of control plants. As a result, WUE was higher for droughted than for control leaves while the CO<sub>2</sub> uptake ratios were quite similar.

#### Magnitude of CAM expression

At 30 days, droughted plants had OA levels and total chlorophyll contents 1.3 and 3.2 times, respectively, lower than control plants (Table 4), however, SI and SOD activity were significantly higher than those for control plants. The higher SOD activity of droughted plants could be related to a higher production of superoxide radicals in response to the increasing magnitude of CAM expression, also could explain their lower leaf chlorophyll content.

In CAM plants such as pineapple, OA levels are closely related with malic acid levels (Chen et al., 2004). The lower OA levels of droughted plants at 6:00 h were likely to be the result of their low  $CO_2$  uptake during night. The reduced chlorophyll content could be the activation of degradation. An increase in the SI has been related to increased succulence (Herrera, 2009). The increase in SI as a result of drought indicates that leaf total chlorophyll decreased more rapidly than did leaf water content.

**Table 4.** Behaviour of variables related with CAM magnitude in micropropagated pineapple plants under water deficit.

Treatments	Organic acid levels at 06:00 h (µeq. g <sup>-1</sup> FW)		Total chlorophyll content at 15:00 h (µg cm <sup>-2</sup> )		Succulence index at 15:00 h (g mg <sup>-1</sup> )		Superoxide dismutase activity at 15:00 h (U mg <sup>-1</sup> Prot.)	
	30 days	45 days	30 days	45 days	30 days	45 days	30 days	45 days
Control plants	133.28 <sup>a</sup>	122.60 <sup>b</sup>	39.32 <sup>b</sup>	46.55 <sup>a</sup>	9.65 <sup>c</sup>	9.42 <sup>c</sup>	$0.021^{b}$	0.015 <sup>c</sup>
Droughted plants	104.17 <sup>c</sup>	124.81 <sup>b</sup>	12.14 <sup>d</sup>	33.54 <sup>c</sup>	15.11 <sup>a</sup>	$12.89^{b}$	$0.053^{a}$	$0.024^{b}$
SE	3.06*		1.77*		0.25*		0.002*	

Different letters and the symbol \* indicates significant differences among the four means within a category. Data were compared using ANOVA factorial followed by Tukey's test ( $p \le 0.05$ ). Analyses were done to n=4 plants, for SOD activity analyses were done to n=3 plants.

The Droughted plants recovered quite rapidly such that after 15 days of being fully watered the OA level was slightly but significantly higher than that of control plants. Total chlorophyll content also recovered and was only about 28% below the level in control plants at 45 days. The SI recovered somewhat but remained higher in droughted plants than in control plants after 45 days because leaf chlorophyll content had recovered relatively more than leaf water content. The SOD activity also recovered, but as with chlorophyll content, it was slower to recover than OA. Elevated SOD activity has been associated with exposure of micropropagated pineapple plants to drought stress during their four first weeks in the acclimatization stage resulted (Aragón et al., 2013), and also was observed when five-month-old micropropagated pineapple plants were exposed to high light intensity at the end of this stage (Rodríguez-Escriba et al., 2015). SOD is one of the primary enzymatic components of the antioxidant system, which converts superoxide into H<sub>2</sub>O<sub>2</sub> and oxygen, it acts like the first defensive response of enzymatic and non-enzymatic mechanisms into cell to remove reactive oxygen species (ROS) (Borland et al., 2006). It is hypothesized that drought-induced SOD activity and other associated physiological changes conditioned plants to better withstand the transition from the acclimatization stage to the field.

#### **CONCLUSION**

Micropropagated pineapple plants of six month old had a stronger CAM performance than did control plants when they were drougthed for 30 days. Their recovery after being fully-watered for 15 days was fast, therefore they can be placed under direct sunlight (high light intensity) for 30 days for their acclimation to this condition and later can be droughted during the same time for induction of defensive mechanism against oxidative stress and excessive water losses. On this way by means of drought after their acclimation to direct sunlight the micropropagated pineapple plants can be hardened to acclimatization-field transition.

#### **ACKNOWLEDGEMENTS**

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#### References

Aragón, C., Carvalho, L., González-Olmedo, J.L., Escalona, M. y Amancio, S. (2012) The physiology of *ex vitro* pineapple (*Ananas comosus* (L.) Merr. var MD-2) as CAM or C3 is regulated by the environmental conditions. Plant Cell Reports 31: 757-769

Aragón, C., Pascual, P., González-Olmedo, J.L., Escalona, M., Carvalho, L. y Amancio, S. (2013) The physiology of *ex vitro* pineapple (*Ananas comosus* (L.) Merr. var MD-2) as CAM or C3 is regulated by the environmental conditions: proteomic and transcriptomic profiles. Plant Cell Reports 32: 1807-1818

- Bartholomew, D. (2009) 'MD2' Pineapple transforms the world's pineapple fresh fruit export industry. Newsletter of Pineapple International Society for Horticultural Sciences 18: 2-5
- Bartholomew, D.P., Paul, R. y Rohrbach, K. (2003) The Pineapple. Botany, production and uses., Ed I Vol 1. CABI, Wallingford
- Borland, A.M., Griffiths, H., Hartwell, J. y Smith, J.A. (2009) Exploiting the potential of plants with Crassulacean Acid Metabolism for bioenergy production on marginal lands. Journal of Experimental Botany 60: 2879-2896
- Escalona, M., Samson, G., Borroto, C. y Desjardins, Y. (2003) Physiology of effects of temporary immersion bioreactors on micropropagated pineapple plantlets. In Vitro Cellular & Developmental Biology Plant 39: 651-656
- FAOSTAT (2013) Food and Agriculture Organization of the United Stated Nations (FAOSTAT). *In* Helping to build a world without hunger,
- Freschi, L., Nievola, C.C., Rodrigues, M.A., Domingues, D.S., Van Sluys, M.A. y Mercier, H. (2009) Thermoperiod affects the diurnal cycle of nitrate reductase expression and activity in pineapple plants by modulating the endogenous levels of cytokinins. Plant Physiology 137: 201-212
- Herrera, A. (2009) Crassulacean Acid Metabolism and fitness under water deficit stress: if not for carbon gain, what is facultative CAM good for? Annals of Botany 103: 645-653
- Lüttge, U. (2004) Ecophysiology of Crassulacean Acid Metabolism (CAM). Annals of Botany 93: 629-652
- Lüttge, U. (2010) Ability of Crassulacean Acid Metabolism plants to overcome interacting stresses in tropical environments. AoB Plants 2010: 1-9
- Nelson, E. y Sage, R.F. (2008) Functional constraints of CAM leaf anatomy: tight cell packing is associated with increased CAM function across a gradient of CAM expression. Journal of Experimental Botany 59: 1841-1850
- Nievola, C.C., Kraus, J.E., Freschi, L., Sousa, B.M. y Mercier, H. (2005) Temperature determines the occurrence of CAM or C3 photosynthesis in pineapple plantlets grown *in vitro*. In Vitro Cellullar and Developmental Biology 41: 832–837
- Prigge, M. y Guriérrez-Soto, M.V. (2014) Pineapple photosynthesis and leaf sap pH as a surrogate of CAM performance in the field. A reseach advance. Pineapple News.21: 18-23. http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews21.pdf
- Rodríguez-Escriba, R.C., Rodríguez, R., López, D., Lorente, G.Y., Pino, Y., Aragón, C.E., Garza-García, Y., Podestá, F.E. y González-Olmedo, J.L. (2015) High Light Intensity Increases the CAM Expression in "MD-2" Micro-Propagated Pineapple Plants at the End of the Acclimatization Stage. American Journal of Plant Sciences 6: 3109-3118
- Rodriguez, R., Aragon, C.E., Escalona, M., Gonzalez-Olmedo, J.L. y Desjardins, Y. (2008) Carbon metabolism in leaves of micropropagated sugarcane during acclimatization phase. In Vitro Cellular & Developmental Biology Plant 44: 533-539
- Rodríguez, R., Becquer, R., Pino, Y., Rodríguez-Escriba, R.C. y López, D. (2013) Introduction of pineapple vitroplantas to field conditions in collaboration with farmers. Preliminary results. Pineapple News 20: 51-56. <a href="http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews20.pdf">http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews20.pdf</a>.

# Response of 'MD-2' Pineapple Plantlets (*Ananas comosus* var. *comosus*) to a Controlled Release Fertilizer During the Acclimatization Stage

Gustavo Y. Lorente González, René C. Rodríguez-Escriba, Roberto E. Izquierdo Martínez, Lucía S. Pérez Borroto, Yuniesky Lobaina Domínguez, Romelio Rodríguez Sánchez, Justo L. González-Olmedo. Agrobiology Laboratory, Bioplants Center. University of Ciego de Avila. Cuba.

#### **Summary**

The pineapple (*Ananas comosus* var. *comosus*) is the second tropical crop of global importance. The variety 'MD-2' is the most commercial and demanded in the United States and Europe. This variety has high yields and good fruit quality, but production of propagules is low, so it takes a long time to get enough plant material when the crop starts. Micro-propagation protocols can produce a large amount of high-quality propagules, but there are still difficulties with the methodology established for acclimatization, especially in the time required for the plants to reach commercial size. The aim of this study was to evaluate the controlled release fertilizer Multicote®, produced by Haifa Chemicals Ltd, to increase the quality of plantlets of pineapple in the acclimatization stage. Two concentrations of Multicote® (0.2 and 0.8 g / plant) and a control were evaluated to determine their effect on morpho-physiological variables. Fertilizer application in mixture with the substrate during the start of the acclimatization stage significantly increased the growth of pineapple plantlets, including significant increases in D-leaf width and length. The CO<sub>2</sub> exchange rate increased with increasing quantity of fertilizer while transpiration decreased with increasing fertilizer. As a result, water use efficiency was much higher in the fertilizer treatments and increased as the amount of fertilizer was increased. It is assumed that higher water use efficiency would improve propagule acclimitization.

## INTRODUCTION

Pineapple is one of the most important fruits in the world and is important for processing and fresh fruit for internal consumption and for exportation (Bartholomew et al., 2003). It is also the most important of CAM plants (Borland et al., 2009). In the last decade the world pineapple production has grown at an average annual rate of 1.9% despite the occurrence of adverse economic and climatic phenomena (FAOSTAT, 2013).

In Cuba the 'MD-2' hybrid was introduced to production scale by Agroindustrial Ceballos Enterprise in 2009 and it is well adapted to the soil and climate conditions of the Ciego de Avila province, so that the production of this commercial variety is rising (Rodriguez et al., 2013). The development of micro-propagation techniques has made the rapid spread of various species of economically important plants possible and pineapple is one of the most studied species. However, despite the advances obtained by micro-propagation, there are still limitations in acclimatization protocols, especially with respect to plant development and the reduction of dwell time of the plantlets (González-Olmedo et al., 2005). At Bioplantas Center the time in the acclimatization stage of Smooth Cayenne pineapple "Serrana" was reduced with the use of weekly foliar fertilization (Yanes et al. (2000), but the production costs increased significantly.

It is therefore necessary to know if the application of the controlled release fertilizer Multicote® during the acclimatization stage of 'MD-2' pineapple plantlets improves their quality and reduces their cost.

## MATERIALS AND METHODS

This research was conducted in specialized laboratories and acclimatization areas of Bioplants Center at the University of Ciego de Avila "Maximo Gomez Baez" (41° 53'N, 78° 41'W, 45 m asl) between December and May. The test material was micro-propagated 'MD-2' pineapple plantlets (*Ananas comosus* var. *comosus*) produced using the protocol by {Daquinta, 1997 #45@@author-year}.

# Effect of Multicote® on the morphology of plantlets during acclimatization

Plantlets selected from plastic culture vessels were about 4.0 cm tall, 7.0 to 8.0 g fresh weight (FW), and had 5 or 6 leaves (Escalona et al., 2003). Plants were dipped in 3.0 ml L-1 of Previcur Energy® (Bayer Crop Science) for 5 min and planted in 250 cm3 plastic containers filled with a 1:1 (v:v) mixture of red ferralytic soil and sugarcane bagasse filter cake (Villalobos et al., 2012). Plants were acclimated into a greenhouse during five months under 80%  $\pm$  3% relative humidity, 25.5°C  $\pm$  2°C temperature and 250  $\pm$  25  $\mu$ mol m-2s-1 photosynthetic

photon flux (PPF). Sprinkler irrigation was done for 30 min at 9:00 am daily. The fertilization treatments were made as follows: Control (foliar application of fertilization every 7 days with COMBI II (5 mL L-1) produced by CARISOMRA SA (Yanes et al., 2000); Multicote® at 0.2 or 0.8 g in 80 cm3 of substrate (5 or 20 granules per well). The dose used was based on recommendations by the manufacturer for use on other crops. After 90 days data on number of leaves, number of roots, plant height (cm), "D" leaf length and width (cm) and plant fresh and dry weight (g) were collected.

#### Effect of Multicote® on physiological variables of pineapple plantlets.

Gas exchange was measured on the middle of the "D" leaves of three plants with a PP Systems, CIRAS-2 Portable Photosynthesis System and PLC6 cuvette (with an analysis area of  $1.7 \text{ cm}^2$ ). The CO<sub>2</sub> exchange (µmol CO<sub>2</sub> m<sup>-2</sup>s<sup>-1</sup>) and transpiration rates (mmol H<sub>2</sub>O m<sup>-2</sup>s<sup>-1</sup>) were measured between 9:00 and 10:00 h at 90 days after treatments were imposed. The equipment was automatically calibrated before each measurement. Instantaneous water use efficiency (WUE) was calculated as transpiration rate (mmol H<sub>2</sub>O m<sup>-2</sup>s<sup>-1</sup>) divided by CO<sub>2</sub> assimilation rate (µmol CO<sub>2</sub> m<sup>-2</sup>s<sup>-1</sup>).

#### RESULTS AND DISCUSSION

Plantlet fresh weights were significantly greater in the Multicote<sup>®</sup> treatments than in the control, but there was no significant difference between Multicote<sup>®</sup> doses (Figure 1). There were no significant differences in the dry weights of plantlets of the three treatments, indicating that the difference in fresh mass could be related to a higher content of water in the tissues of plants in the Multicote<sup>®</sup> treatments.

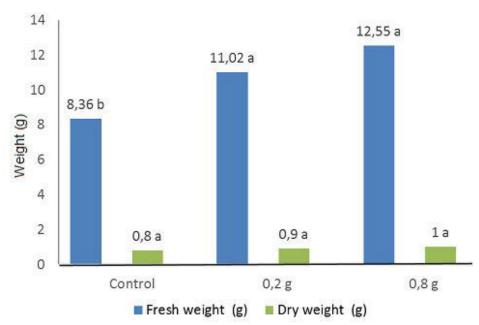


Figure 1. Effect of applying Multicote® on fresh and dry weight of pineapple plantlets during the acclimatization stage. Means with different letters indicate significance by one way ANOVA and the Tukey multiple range test, p < 0.05). Each data point represents the mean for n=20.

The increase in fresh mass in the Multicote<sup>®</sup> treatments resulted in significant increases over the control in all morphological variables (Table 1). The lengths and widths of the D leaves increased significantly as the amount of Multicote<sup>®</sup> was increased. The use of Multicote<sup>®</sup> clearly increased growth relative to weekly foliar fertilization.

Table 1. Effect of Multicote ® on morphological variables in pineapple during acclimation.

Treatment	Leaves number	Roots number	Plant height	D-Leaf length (cm)	D Leaf width (cm)	Length / width of D Leaf
Control	11.8 b <sup>†</sup>	9.0 b	10.8 b	7.5 c	1.7 c	4.4 b
0.2 g	13.0 a	11.0 a	12.5 a	10.4 b	1.9 b	5.4 a
0.8 g	13.3 a	10.8 a	14.6 a	11.5 a	2.1 a	5.4 a
$SE(\overline{X})$	0.32	0.62	0.68	0, 2	0.09	0.11

<sup>&</sup>lt;sup>†</sup>Data within a column followed by the same letter are not significantly different from each other based on one way ANOVA and the Tukey multiple range test (p <0.05). Each data point represents the mean for n = 20.

Leaf CO<sub>2</sub> exchange rate was lowest in the control and increased as the dose of Multicote<sup>®</sup> increased (Table 2). On the other hand plants fertilized with 0.8 g of Multicote<sup>®</sup> transpired less, indicative of improved water control and much greater water use efficiency, presumably because of improved stomatal functionality.

Table 2. Effect of Multicote<sup>®</sup> on the photosynthetic activity, transpiration and efficiency in water use of 'MD-2' pineapple plantlets.

Teatments	$CO_2$ exchange rate. ( $\mu$ mol $CO_2$ m <sup>2</sup> s <sup>-1</sup> )	Transpiration rate (mmol (H <sub>2</sub> O) m <sup>2</sup> s <sup>-1</sup> )	Nater Use Efficiency (μmol CO <sub>2</sub> /mmol H <sub>2</sub> O)
Control	2.86 c	0.31	9,22 c
0.2 g	3.20 b	0.23 b	13.91 b
0.8 g	4.95 a	0.15 c	33,00 a

It has been observed that transpiration and CO<sub>2</sub> assimilation rate in pineapple plantlets grown in a greenhouse are very sensitive to light intensity and relativity humidity {Rodríguez-Escriba, #43;Rodríguez-Escriba, 2015 #1568}. The greater water use efficiency (WUE) obtained with the highest dose of Multicote<sup>®</sup> fertilizer would indicate that the use of a controlled release fertilizer could improve the acclimatization of plantlets.

#### **CONCLUSIONS**

Doses of 0.2 and 0.8 g of Multicote<sup>®</sup> fertilizer mixed with the substrate from the start of the acclimatization stage stimulated all morpho-physiological variables evaluated in pineapple plantlets and this enhanced their commercial quality. The greatest response was to 0.8 g per plant of Multicote<sup>®</sup> fertilizer. That treatment stimulated the  $CO_2$  exchange rate and reduced seedling transpiration, which greatly increased water use efficiency, which relative to other treatments is assumed to improve adaptation at this development stage.

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#### References

Bartholomew, D. P., Paull, R. E. & Rohrbach, K. G. 2003. The pineapple: botany, production, and uses, Wallingford, UK, CABI.

Borland, A. M., Griffiths, H., Hartwell, J. & Smith, J. A. 2009. Exploiting the potential of plants with Crassulacean Acid Metabolism for bioenergy production on marginal lands. J Exp Bot, 60, 2879-96.

Daquinta, M. & Benegas, R. 1997. Brief review of tissue culture of pineapple. Pineapple News, 3, 7-9.

Escalona, M., Samson, G., Borroto, C. & Desjardins, Y. 2003. Physiology of effects of temporary immersion bioreactors on micropropagated pineapple plantlets. In Vitro Cellular & Developmental Biology - Plant, 39, 651-656.

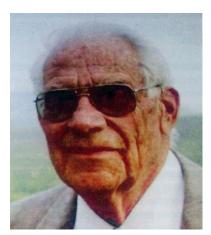
FAOSTAT 2013. Helping to build a world without hunger, Roma, Food and agriculture organization of the United Stated Nations (FAOSTAT).

- González-Olmedo, J. L., Fundora, Z., Molina, L. A., Abdulnour, J., Desjardins, Y. & Escalona, M. 2005. New contributions to propagation of pineapple (Ananas comosus L. Merr) in temporary immersion bioreactors. In Vitro Cellular & Developmental Biology Plant, 41, 87-90.
- Rodríguez-Escriba, R., López, D., Aragón, C., Becquer, R., Pino, Y., Garza, Y., Podestá, F. E., González-Olmedo, J. L. & Rodríguez, R. 2014. CAM Metabolic Changes of 'MD-2'Pineapple Grown Under High and Low Light. Newsletter of the Pineapple Working Group, International Society for Horticultural Science, 21, 27-34.
- Rodríguez-Escriba, R. C., Rodríguez, R., López, D., Lorente, G. Y., Pino, Y., Aragón, C. E., Garza-García, Y., Podestá, F. E. & González-Olmedo, J. L. 2015. High Light Intensity Increases the CAM Expression in "MD-2" Micro-Propagated Pineapple Plants at the End of the Acclimatization Stage. American Journal of Plant Sciences, 6, 3109-3118.
- Rodríguez, R., Becque, R., Pino, Y., Rodríguez, R.C., Lopez, D. & González, J.L. 2013. Introduction of Pineapple Vitroplantas to Field Conditions In Collaboration with Farmers. Preliminary Results. Pineapple News, No. 20, 51-56.
- Rodríguez, R., Becque, R., Pino, Y., Rodríguez, R.C., Lopez, D. & González, J.L. 2013. Introduction of Pineapple Vitroplantas to Field Conditions In Collaboration with Farmers. Preliminary Results. Pineapple News, No. 20, 51-56.
- Villalobos, A., González-Olmedo, J., Santos, R. & Rodríguez, R. 2012. Morpho-physiological changes in pineapple plantlets [Ananas comosus (L.) Merr.] during acclimatization. Ciência e Agrotecnologia, 36, 624-630
- Yanes, P., González, O. & sánchez, R. 2000. A technology of acclimatization of pineapple vitroplants. Pineapple News, 7, 24.

# **News from France**

# Claude Py, in Memorium

Jean Marchal and Claude Teisson, CIRAD (retired)



Dr. Claude Py

Claude Py was born in 1923 in north-eastern France, he passed away on July 29, 2015. He was a pioneer and a world-renowned researcher in the agronomy of pineapple, to which he devoted his entire caeer. He made major contributions to the knowledge and cultivation of this species.

After graduating as an agronomist at Grignon Agronomical Engineering School (France), he specialized in tropical agronomy and genetics at ORSTOM (Office of Scientific and Technical Research Overseas, which later became the IRD (French Research Institute for Development). Subsequently, he spent his entire career with the IFAC (French Fruit and Citrus Institute), which was established in 1976 at CIRAD (French Centre for International Cooperation in Agricultural Research for Development).

His energy, enthusiasm, humor, optimism and tolerance were personality traits that were appreciated by everyone who knew him, friends and colleagues alike. They were probably influenced by the ordeal of his deportation to Dachau concentration camp in Germany (1944-1945) and his

almost miraculous survival.

After a fact-finding mission on pineapple in the United States and in the West Indies, in 1950, he was assigned to the IFAC research center in Conakry, Guinea ,then to the French West Indies, and ended his career in Mainland France. He retired in 1985, but maintained his interest in pineapple research and continued to play an active role in community life in Montpellier.

Throughout his career and as program director at IFAC and CIRAD, he surrounded himself with an experienced team in each of the different fields of research and development in which he was involved. He worked for international organizations including the World Bank, the FAO, the European Community, as well as for State enterprises and private firms as a consultant and as an expert on the quality of research, and on opportunities for the further development of pineapple. As such, he visited and gave advice in the majority of pineapple producing countries or regions with pineapple production potential.

He helped to establish relationships and set up joint work programs with a number of scientific organizations including the University of Hawaii, Embrapa and the University of Campinas in Brazil, the Ministry of Research and Agronomic Institute in Cuba and many official bodies in Asia, America and especially in Africa to which he was particularly attached.

C. Py published a great number scientific articles, many in the scientific journal Fruits. He also wrote articles for the general public, expert reports, and provided advice and recommendations for contracting agencies as well as writing internal documents outlining ongoing research and research results.

His first book "Pineapple" (C. Py and MA. Tisseau) was published in 1965. His second book "Pineapple, its cultivation, its products" (C. Py, J-J. Lacoeuilhe, C. Teisson) was published in 1984. An updated English version "The pineapple, cultivation and uses" was published in 1987. This is still the most comprehensive reference work on the crop published to date. The book has been out-of-print for many years; however, some new and used copies can still be found for sale at Amazon.com.

His human qualities made him a great team leader, a true "master" in the old sense of the word, transmitting not only scientific knowledge but human values.

# A Tribute from EMBRAPA

Dr. Claude Py gave great contributions to the pineapple research program at Embrapa Cassava & Fruits, Cruz das Almas, Bahia, Brazil, set up in the late seventies and early eighties. His large experience and easy communication allowed Embrapa's pineapple research team to learn a lot during his visits to Brazil, as well as during training

activities carried out at the laboratories in Montpellier and Cirad experimental fields in some African countries, especially Côte d'Ivoire, which was one of the main pineapple exporters at that time. For a long time his books have been our main consultation tool on pineapple cultivation. The partnership Cirad – Embrapa, constructed under his leadership, has been continued for decades based upon excellent scientific interactions on all aspects of pineapple research. In the name of all Embrapa colleagues that have had the satisfaction and the benefits of this overseas alliance we express our gratefulness and admiration.

Domingo Haroldo Reinhardt and Aristoteles Pires de Matos Embrapa Cassava & Fruits, Cruz das Almas, Bahia, Brazil

# **Claude Py Publications**

(Paste titles in French into <a href="https://translate.google.com/">https://translate.google.com/</a> for a translation.)

Aubert, B., Gaillard, J.P., Lossois, P., Py, C., and Marchal, J., 1973. Influence de l'altutude sur le comportement de l'ananas "Cayenne Lisse". Essais realises au pied du mont Cameroun. Fruits 28:203-214.

Barbier, M. and Py, C., 1958. Nouveaux progres dans la culture de l'ananas a Porto-Rico. Fruits 13:15-20.

Brun, J. and Py, C., 1952. Leaf symptoms of zinc deficiency in pineapple in French Guinea. Fruits 7:62-64.

Giacomelli, E.J. and Py, C., 1981. O abacaxi no Brasil. Fundação Cargill, Campinas.

Giacomelli, E.J. and Py, C., 1981. L'ananas au Bresil. Fruits 36:645-687.

Giacomelli, E.J. and Py, C., 1982. Expansão da abacaxicultura no Brasil. Fundação Cargill, Campinas.

Giacomelli, E.J., Py, C., and Lossois, P., 1984. Estudo sobre o ciclo natural do abacaxizeiro 'Cayenne' no planalto paulista. Bragantia Campinas 43:629-642.

Godefroy, J., Py, C., and Tisseau, M.A., 1971. Action de la fumure phosphatee en culture d'ananas en Cote d'Ivoire et en Guadeloupe (Effect of phosphate fertilizer on pineapple in the Ivory Coast and in Guadeloupe). Fruits 26:207-210.

Guyot, A., Pinon, A., and Py, C., 1974. The pineapple in the Ivory Coast. Fruits 29:85-117.

Guyot, A. and Py, C., 1970. La floraison controlee de l'ananas par l'ethrel, nouveau regulateur de croissance (2eme partie). Fruits 25:341-347.

Guyot, A. and Py, C., 1970. La floraison controlee de l'ananas par l'ethrel, nouveau regulateur de croissance. Fruits 25:427-445.

Lacoeuilhe, J.J. and Py, C., 1974. La croissance de la feuille d'ananas en Cote d'Ivoire. Fruits 29:709-715.

Py, C., 1949. The classification, origin, distribution and genetics of the pineapple. Fruits 4:407-414.

Py, C., 1949. Fasciation in pineapple. Fruits 4:180-182.

Py, C., 1950. The classification, origin, distribution and genetics of the pineapple. Fruits 5:5-12.

Py, C., 1950. l'ananas aux iles Hawaii. Fruits 5:124-133.

Py, C., 1951. An attempt to improve pineapple culture in French Guinea. Fruits 6:235-237.

Py, C., 1952. La reduction de la couronne d'ananas. Fruits 7:392-398.

Py, C., 1952. New information on fasciation in pineapples. Fruits 7:342-346.

Py, C., 1952. Note on pineapple cultivation in the Ivory Coast. Fruits 7:195.

Py, C. 1952. Nouvelles données sur les traitements hormones.

Py, C., 1952. Pineapples in Cuba. Fruits 7:180-182.

Py, C., 1953. Les hormones dans la culture de l'ananas. Ann Inst Fruits Agrumes Colon 6:46.

Py, C., 1954. Le probleme de las lutte contre les mauvaises herbes dans les plantations d'ananas. Fruits 9:191-202.

Py, C., 1955. Los differents types de rejets d'ananas. Fruits 10:25-42.

Py, C., 1955. The effect of manuring on the response of pineapples to acetylene treatment. Rev Agric Reunion 55:88-91.

Py, C., 1955. Le CMU herbicide selectif. hautement effrace pour plantation d'ananas. Fruits d'Outre Mer. 10:157-161.

Py, C., 1955. Stude du prix de revient de l'emballage des ananas. Fruits 10(9):389-394.

Py, C., 1955. Variation, chez l'ananas, de l'ecart de temps qui separe le traitement destine a provoquer la floraison de la recolte. Rev Agric Reunion 55:85-88.

Py, C., 1957. Recherche d'une methode de lutte economique contre l'Imperata cylindrica dans les plantations d'ananas. Fruits 12:377-384.

Py, C., 1958. La production d'ananas de novembre-decembre en Guinea. Fruits 13:105-110.

Py, C., 1958. Packing pineapples. Fruits 13:67-71.

Py, C., 1959. Etude sur la croissance de l'ananas en Guinee. Fruits 14:3-24.

Py, C., 1959. Experiments in pineapple packing, season 1958-1959. Fruits 14:467-470.

Py, C., 1959. La lutte contre les mauvaises herbes en plantation d'ananas. Fruits 14:247-261, 291-329.

Py, C., 1960. The effects of planting date and sucker weight on the growth of pineapple plants in Guinea. Fruits 15:451-453.

Py, C., 1960. The storage of pineapple suckers during the dry season. Fruits 15:29-32.

# Newsletter, Pineapple Working Group, International Society for Horticultural Science

- Py, C., 1961. L'influence des facteurs climatiques sur l'efficacite de la fumure azotee en plantation d'ananas. Fruits 16:375-378.
- Py, C., 1961. Quelques tests sur la floraison "provoquee" de l'ananas. Fruits 16:28.
- Py, C., 1962. Comparaison de differents selections d'ananas Cayenne lisse et de plusieurs outres varietes. Fruits 17(11):559-572.
- Py, C., 1962. Comparaison de l'uree et du sulfate d'ammoniaque pour la fumure de l'ananas en Guinee. Fruits 17:95-97.
- Py, C., 1962. A comparison of different selections of Smooth Cayenne and various other pineapple varieties. Fruits 17:559-572.
- Py, C., 1962. Foliar spraying with urea as applied to pineapples. Fruits 17:285-287.
- Py, C., 1962. Techniques de production et d'expedittion des ananas frais, produc guadel sucr. Fruits 4:2-6.
- Py, C., 1963. Determination of the best times for fertilizer application to pineapples. Results of experiments begun in Guinea in 1959 on urea application during fruiting. Fruits 18:75-77.
- Py, C., 1964. Apercu sur le cycle de l'ananas en Martinique. Fruits 19:133-139.
- Py, C., 1964. Pineapple fertilization. Is sulphur responsible for the superiority of ammonium sulphate over urea? Fruits 19:529-535.
- Py, C., 1965. Approches pour combler le deficit en eau, principal facteur limitant de la culture de l'ananas en Guinee. Fruits 20:315-329.
- Py, C., 1965. Comparative study of the pineapple industries of hawaii, formosa, the philippines and malaysia 1.hawaii 2.formosa. Fruits 20:59-70,99-107.
- Py, C., 1965. Comparative study of the pineapple industries of hawaii, formosa, the philippines and malaysia 3.philippines 4. Malaysia. Fruits 20:141-150.
- Py, C., 1968. Chemical weed control in pineapple plantations in the light of recent experimental results. Fruits 23:3-12.
- Py, C., 1968. Interet, dans la culture de l'ananas en zone humide, d'une couverture du sol en polyethylene. Fruits 23:139-148.
- Py, C., 1969. Mechanization problems in pineapple cultivation. Fruits 24(1):5-19.
- Py, C., 1970. The best dates for planting pineapples intended for processing in martinique. Fruits 25:199-203.
- Py, C., 1970. Controlled maturation of pineapples. Fruits 25:693-694.
- Py, C., 1972. An outline of the state of pineapple research. Fruits 27(2):107-109.
- Py, C., 1973. Culture techniques, principal problems, solutions looked upon for resolutions. Gaz Agric (Luanda) 18(5):319,321,332.
- Py, C., 1973. The pineapple, its quality and cost in relation to the environment. Fruits 28(2):127-131.
- Py, C., 1973. Pineapples in Angola. Gaz Agric (Luanda) 18(9):648-649.
- Py, C., 1975. Etude comparee des industries de l'ananas aux iles Hawai,a Formose, aux Philippines et en Malaysis. Fruits 20(3):99-107.
- Py, C., 1976. Study and development of pineapple (culture) in West Africa, especially on the Ivory Coast. Acta Horticulturae 57:11-120.
- Py, C., 1979. Accelerated production of vegetative planting material (pineapples). Fruits 34:107-116.
- Py, C., 1979. The latest on weed control in pineapples. Compte Rendu De La 10th Conference Du Columa:1136-1144.
- Py, C., 1981. Economic benefit of herbicides in village pineapple crops. Compte Rendu De La 11 Conference Du Columa,Irfa/Gerdat 3:933-940.
- Py, C., 1985. l'ananas au perou. Fruits 40(2):97-111.
- Py, C. and Al, E., 1957. Fertilizing pineapples in Guinea. Fertilite 3:5-25.
- Py, C. and Barbier, M., 1953. The production of pineapple in French Guinea for export as fresh fruit. Fruits 8:363-392.
- Py, C. and Barbier, M., 1966. New pineapple culture techniques in the west indies. Fruits 21:229-230.
- Py, C. and Estanove, P., 1964. La multiplication des ananas par portions de tige. Es'tude de que lques facteurs susceptibles den influencer les resultats. Fruits 19:465-468.
- Pv, C. and Fouque, A., 1963. Fruit crops of Puerto Rico. Fruits 18:325-336.
- Py, C. and Fouque, A., 1964. La fumure de l'ananas en Guyane. Fruits 19:262-264.
- Py, C. and Gaillard, J.P., 1971. La formation el la croissance des rejets d'ananas. Fruits 26:211-222.
- Py, C., Giacomelli, G., and Lossois, P., 1973. Essai densite, conduit a caprim verde (Sao Paulo, Bresil) pour le compte de citrobrasil. Reunion Annuelle Ifac Doc 45:3P.
- Py, C. and Guyot, A., 1970. La floraison controlee de l'ananas par l'ethrel, nouveau regulator de croissance (l'ere partie). Fruits 25:253-262.
- Py, C. and Guyot, A., 1970. Study on the utilization of pineapple in the cannery. Fruits 25:349-356.
- Py, C. and Haendler, L., 1952. Utilisation des dechets de conserveries d'ananas. Fruits 7(5):231-233.
- Py, C., Haendler, L., Huet, R., and Silvy, A., 1956. La fumure de l ananas en Guinee. Fruits 11:5-23.
- Py, C., Lacoeuilhe, J.J., and Teisson, C., 1987. The pineapple. Cultivation and uses. Editions G.-P. Maisonneuve, Paris.
- Py, C. and Lossois, P., 1962. Prevision de recolte en culture d'ananas. Etudes de correlations. Deuxieme partie. Fruits 17:75-87.

## Newsletter, Pineapple Working Group, International Society for Horticultural Science

- Py, C., Lossois, P., and Karamkam, M., 1968. Contribution to the study of the growth cycle of pineapples. Fruits 23:403-413.
- Py, C. and Naville, R., 1973. Economic production of quality pineapples. Shell Publ Helath Agric News 16(1):21-23.
- Py, C. and Others, 1958. Effect of mineral elements on the composition of pineapples. Qual Plant Mayeg 3/4:237-243.
- Py, C. and Pelegrin, P., 1958. Prevision de recolte en culture d'ananas. Fruits 13:243-251.
- Py, C. and Silvy, A., 1954. Traitements hormones sur ananas. Methode pratique pour diriger la production. Fruits 9:101-123.
- Py, C. and Tisseau, M.A., 1965. L'ananas. G.-P. Maisonneuve et Larose, Paris.
- Py, C., Tisseau, M.A., Oury, B., and Ahmada, F., 1957. La culture de l'ananas en Guinee--Manuel du planteur. Institute Français De Recherches Fruitieres D'outre-Mer (I.F.A.C.), Paris, Françe.
- Teisson, C., Martin-Prevel, P., Combres, J.C., and Py, C., 1978. Internal browning of pineapple, a disorder caused by refrigeration. Fruits 33(1):48-50.

# The Last Revision of Pineapple Nomenclature

Geo Coppens d'Eeckenbrugge CIRAD, UMR AGAP, Avenue Agropolis, 34398 Montpellier Cedex 5

The revision of synonymies in pineapple taxonomy, announced in the last issue of our newsletter (Coppens d'Eeckenbrugge, 2015), has been completed and published (Coppens d'Eeckenbrugge and Govaerts, 2015). This publication does not alter the current classification (Coppens d'Eeckenbrugge and Leal, 2003), except for one name. Thus, for those researching any aspect of pineapple cultivated for its fruit the name of the plant is *Ananas comosus* var. *comosus*. If the author name is required, as for example in the title of a scientific article, it should be written *Ananas comosus* (L.) Mill. var. *comosus*.

From a taxonomic point of view, the first practical consequence of this formal work is the restoration of *Ananas comosus* var. *microstachys* (Mez) Smith instead of *Ananas comosus* var. *ananassoides* (Baker) Coppens & F.Leal. A second consequence is that our classification, and the associated synonymies, is now fully recognized in the World Checklist of Selected Plant Families (WCSP) (<a href="http://apps.kew.org/wcsp/qsearch.do">http://apps.kew.org/wcsp/qsearch.do</a>), which should widen its acceptance well beyond the circle of plant breeders and germplasm curators.

A search on the WCSP website currently yields as many as 88 synonyms for our two species and five botanical varieties. Most of them are anecdotal, and after revision we can bury them back in our memory, as their interpretation does not interfere appreciably with the modern classification. Others correspond to more important steps in the evolution of pineapple nomenclature. Below are listed the most important synonyms recognized by Coppens d'Eeckenbrugge and Leal (2003) and Coppens d'Eeckenbrugge and Govaert (2015).

## Ananas macrodontes Morren (1878) (modern)

This is the *yvira*, a wild tetraploid pineapple without crown, propagated vegetatively by stolons instead of suckers. Key synonyms: *Pseudananas macrodontes* (Morren) Harms (1930), *Pseudananas sagenarius* (Arruda da Câmara) Camargo (1939).

Ananas comosus (L.) Merrill (1917) (modern, as are the botanical varieties listed below)

This diploid species includes the following five botanical varieties  $(1-5\ below)$ , three of which are domesticates.

# (1) Ananas comosus var. comosus (the edible pineapple)

This is the edible pineapple, known as *Ananas comosus* from 1917 to 2003, then classified more precisely as a botanical variety, to accommodate four other, wild and cultivated, forms at the same rank in the same species. Key synonyms: *Ananas sativus* Schultes & Schultes (1830; spiny cultivars), *Ananas semiserratus* (Willd.) Schultes & Schultes (1830; cultivars with spines at leaf apex), *Ananas lucidus* (Aiton) Schultes & Schultes (1830; smooth-leaved cultivars), *Ananas debilis* (Lindley) Schultes & Schultes (1830; a lost wave-leaved cultivar), *Ananas comosus* f. *sativus* (Schult. & Schult.f.) Mez (1934), *Ananas comosus* f. *lucidus* Mez (1934).

#### (2) Ananas comosus var. microstachys (Mez) Smith (1934)

This is the most common wild pineapple, with long and narrow leaves and a small fruit, from which the edible pineapple was domesticated. It is distributed in neotropical areas east of the Andes, under conditions of climatic and/or edaphic drought (rocks and sandy soils).

Key synonyms: Acanthostachys ananassoides Baker (1889), Ananas microstachys Lindman (1891), Ananas sativus Schult. & Schult.f. var. microstachys Mez (1892), Ananas ananassoides (Baker) Smith (1939), Ananas comosus var. ananassoides (Baker) Coppens & F.Leal (2003), Ananas ananassoides Baker var. nanus Smith (1939), Ananas microstachys var. nanus (L.B.Sm.) Camargo (1942), Ananas nanus (L.B.Sm.) Smith (1962).

#### (3) Ananas comosus var. parguazensis (Camargo & L.B.Sm.) Coppens & Leal (2003)

Another wild botanical variety of pineapple, with wider leaves and some retrorse spines, mostly found in the basins of Rio Orinoco (south-eastern Colombia and southern Venezuela) and Rio Negro (north-western Brazil), and more rarely in the Guianas, together with *Ananas comosus* var. *microstachys*, and forms that appear intermediate between the two wild botanical varieties.

Key synonym: Ananas parguazensis Camargo & Smith (1968)

## (4) Ananas comosus var. erectifolius (L.B.Smith) Coppens & Leal (2003)

This is the *curagua*, a small-fruited pineapple, cultivated only for fiber (Leal & Amaya, 1991), north of the Amazon River (Guianas and Venezuela). This cultigen (known in cultivation only) evolved from *Ananas comosus* var. *microstachys* through multiple domestication events (Duval et al. 2003), which determined its most characteristic features: erect leaves, related to selection for a high fiber content, and absence of marginal spines, related to a dominant mutation (Collins 1960).

Key synonym: *Ananas erectifolius* Smith (1939). The synonymy with *Ananas lucidus* Miller, a large-fruited and smooth-leaved cultivar, proposed by Smith and Downs (1979) is not founded.

# (5) Ananas comosus var. bracteatus (Lindley) Coppens & Leal (2003)

This cultivated botanical variety is particular in resulting from the introgression of *A. macrodontes* genes into *A. comosus*. It includes two forms that have been propagated vegetatively, which explains their very low genetic diversity. The very rare form, corresponding to *A. fritzmuelleri* Camargo, shares nuclear and cytoplasmic genes with *A. macrodontes*, as well as more morphological traits (longer bracts, retrorse spines). The second form, quite common as an ornamental in tropical gardens, has been given the specific epithets or botanical variety names *bracteatus* (referring to its long bracts) and *sagenaria* (from the Latin word for net, referring to its ancient use as a fiber plant). It appears to share a lesser proportion of nuclear genes and no cytoplasmic genes with *A. macrodontes* (Duval et al. 2001, 2003).

Key synonyms: *Bromelia sagenaria* Arruda da Câmara (1810). *Ananas sagenaria* (Arruda da Câmara) Schultes & Schultes (1830). *Bromelia silvestris* Vellozo (1829). *Ananas silvestris* Müller (1896). *Ananas fritzmuelleri* Camargo (1943), *Ananassa bracteata* Lindley (1827). *Ananas bracteatus* (Lindley) Schultes & Schultes (1830), *Ananas sativus* var. *bracteatus* (Lindley) Mez (1892).

For readers interested in understanding better the history of these synonyms and the evolution of pineapple taxonomy, I present hereafter a summary inspired from the paper of Leal et al. (1998).

Pineapple taxonomy long focused on the description of variation among clones cultivated for the fruit. Indeed, from the late 18<sup>th</sup> century, pineapple was mostly known from cultivation in European glasshouses, and most cultivars of *A. comosus* var. *comosus* were first given Latin names, generating much confusion with the Latin binomials used for species in the Linnaean system. Botanical knowledge of wild forms was very limited. Beer (1856) examined a herbarium specimen of a small-fruited wild type ("a botanical rarity") and concluded that the differences with the cultivated pineapples (hypertrophy of the syncarp) were only the result of cultivation. Further in the same document, he gave more importance to smaller differences observed among groups of cultivars, raising some of them to the species rank. In 1879, when Morren described a second species of pineapple, *Ananas macrodontes*, from glasshouse plants, he ignored its ecology. Finally, Baker (1889) was the first to give a species rank to a wild pineapple. However, he classified it in the genus *Acantostachys* (*A. ananassoides*), an error which was half-corrected by Lindman in 1891, when the latter classified it as *Ananas microstachys* (in fact, he should have maintained the epithet and named it *Ananas ananassoides*).

Mez (1892) proposed a first simplification, downgrading the common cultivar groups to botanical varieties of a unique species, *Ananas sativus*, and considered the wild pineapple as another botanical variety, *A. sativus* var. *microstachys*. Mez (1892) included *A. macrodontes* within *A. sativus* var. *bracteatus*, because he supposed that the absence of a crown in Morren's description was only the result of observing a juvenile inflorescence. In 1919, two years after Merrill established the binomial *Ananas comosus*, Bertoni (1919) took an opposite direction and divided *Ananas* into five species with many botanical varieties, producing a very confused classification that was never used later. In 1930, Harms created a new, monotypic genus, raising again Morren's *A. macrodontes* to the species rank, under the binomial *Pseudananas macrodontes*.

In 1934, Mez maintained his parsimonious vision, with one genus and only three species. Within *A. comosus*, he included cultivated pineapples as simple "forms" (not botanical varieties), based on leaf spininess and shape. In addition, he recognized *A. macrodontes*, and retained *A. sagenaria* instead of his *A. sativus* var. *bracteatus* of 1892. Surprisingly, he retained no particular status for the wild representatives of *A. comosus*. However, their botanical variety rank was restored by Smith (1934), who proposed *Ananas comosus* var. *microstachys*.

After Mez, pineapple classification returned to a more complex system, as Smith (1939, 1961, 1962, 1971), together with Camargo (1939, 1942, 1943; Camargo and Smith, 1968), multiplied species and botanical varieties without describing significant new variation (except for the re-discovery of the curagua). Many varieties were raised to the species rank, and the genus *Pseudananas* was restored, a process which culminated in a list of two genera and eight species (Smith and Downs, 1979).

The classification of Coppens d'Eeckenbrugge and Leal (2003) may be compared with the parsimonious views of Mez. Although Mez's treatment varied from 1892 to 1934, he mostly considered differences among wild and cultivated crowned pineapples, selected or maintained for distinct purposes (fruit, fiber, ornamental), at the infraspecific level. The similitude is clearer when one compares the synonymies given by Mez and those given in Coppens d'Eeckenbrugge and Leal (2003) and Coppens d'Eeckenbrugge and Govaert (2015). Mez lacked our modern knowledge based on direct observations in the wild, in living collections, and in the field, as well as more data from reproductive biology and molecular genetics, but his acute and critical reading of all the literature then available allowed him to comprehend the essence of pineapple diversity.

#### References

Baker, J.G. 1889. Handbook of the Bromeliaceae. George Bell & Sons, London, 22–25.

Bertoni, M.S. 1919. Contribution à l'étude botanique des plantes cultivées. I. Essai d'une monographie du genre *Ananas*. Anales Científicos Paraguayos (Serie II) 4: 250–322.

Camargo, F.C. 1939. Ananás e abacaxí. Revista de Agricultura (Piracicaba) 14: 331–338.

Camargo, F.C. 1942. Pesquisas taxonômicas sôbre os gêneros *Pseudananas* e *Ananas*. Proceedings of the Eighth American Scientific Congress 3: 178–192.

Camargo, F.C. 1943. Vida e utilidade das Bromeliáceas. Boletim Técnico Instituto Agronômico do Norte 1: 1–31.

Camargo, F.C. and Smith, L.B. 1968. A new species of Ananas from Venezuela. Phytologia 16: 464–465.

Collins, J.L. 1960. The pineapple, botany, utilization, cultivation. Leonard Hill, London.

Coppens d'Eeckenbrugge, G. 2015. A Short Update on the Pineapple Taxonomy Debate. Pineapple News 22: 28.

Coppens d'Eeckenbrugge, G. and Govaerts, R. 2015. Synonymies in *Ananas* (Bromeliaceae). Phytotaxa 239 (3): 273–279.

Coppens d'Eeckenbrugge, G. and Leal, F. 2003. Morphology, anatomy and taxonomy. In: Bartholomew, D.P., Paull. R.E., and Rohrbach, K.G. (Eds.) The pineapple: botany, production and uses. CABI, Wallingford, Oxford, UK. 13–32.

Duval, M-F., Noyer, J-L., Perrier, X., Coppens d'Eeckenbrugge, G., and Hamon, P. 2001. Molecular diversity in pineapple assessed by RFLP markers. Theor. Appl. Gen 102(1): 83-90.

Duval, M-F., Buso, G.C., Ferreira, F.R., Bianchetti, L.B., Coppens d'Eeckenbrugge, G., Hamon, P., and Ferreira, M.E. 2003. Relationships in *Ananas* and other related genera using chloroplast DNA restriction site variation. Genome 46: 990–1004.

Harms, H. 1930. Bromeliaceae. Die natürlichen Pflanzenfamilien 15a: 153.

Leal, F. and Amaya, M-L. (1991) The curagua (*Ananas lucidus*, Bromeliaceae) crop in Venezuela. Economic Botany 45: 216–224.

Leal, F., Coppens d'Eeckenbrugge, G., and Holst, B. 1998. Taxonomy of the genera *Ananas* and *Pseudananas* - An historical review. Selbyana, 19: 227–235.

Lindley, J. 1827. Edwards's Botanical Register. Vol. 13. John Ridgway & Sons, London, pp. 1069 & 1081.

Lindman, C.A.M. 1891. Bromeliaceae. Herbarii Regnellianii. I. Bromeliae. Kongl. Svenska Vetenskaps Academiens Handlingar 24 (8): 1–49.

Merril, E.D. 1917. An interpretation of Rumphius's Herbarium Amboinense. Manila Bureau of Printing, pp. 46, 133–134.

Mez, C. 1892. Bromeliaceae; *Ananas*. Martius, Flora Brasiliensis 3(3). Reprinted 1965 Verlag von J. Cramer, Weinheim, Codicote (Hertfordshire), Wheldon & Wesley, New York, pp. 288–294.

Mez, C. 1934. Das Pflanzenreich. Bromeliaceae. Ananas Adans. Engler Prantl, 160. Reprinted 1965 Verlag von J. Cramer, Weinheim, pp. 101–104.

Morren, E. 1878. Description de l'*Ananas macrodontes* sp. nov. Ananas à fortes épines. La Belgique Horticole - Annales d'Horticulture 28: 140–172.

Müller, F. 1896) Die Bromelia silvestris der Flora fluminensis. Berichte der Deutschen Botanischen Gesellschaft 14: 3–11.

# Newsletter, Pineapple Working Group, International Society for Horticultural Science

- Schultes, J.A. and Schultes, J.H. 1830. Systema Vegetabilium 7: 1283–1287.
- Smith, L.B. 1934. IV. Studies in the Bromeliaceae. Contributions from the Gray Herbarium 104: 72.
- Smith, L.B. 1939. Notes on the taxonomy of *Ananas* and *Pseudoananas*. Botanical Museum Leaflets Harvard University 7: 73–81.
- Smith, L.B. 1961. Notes on Bromeliaceae, XVII. Phytologia 8 (1): 1–13.
- Smith, L.B. (1962) A new look at the species of pineapple. Bulletin of the Bromeliad Society 12: 54–55.
- Smith, L.B. and Downs, R.J. .1979. Bromelioideae (Bromeliaceae). Flora Neotropica, Monograph 14(3). New York Botanical Garden, New York, pp. 1493–2142.

# **News from the USA (Hawaii)**

# Solar Injury Causes Crown Deformities of 'CO-2' Fruits

Duane P. Bartholomew and Glenn Taniguchi duaneb@hawaii.edu; gtani@hawaii.edu

Various crown abnormalities have been observed in Australia, Hawaii and South Africa (Collins, 1950; Newett and Rigden, 2015; Py et al., 1987) and Bartholomew et al (2003) concluded that the principle cause was heat injury to crown apical tissues. Newett and Rigden (2015) state that the incidence of crown injury is greatest when air temperature exceeds 30 °C and there is little or no wind. Py et al. (1987) report that the incidence of crown injury was higher when floral initiation occurred during a dry, sunny period and irrigation, method not specified but assumed to be overhead, reduced the number of affected fruits. The incidence of crown injury is higher in low than in high density plantings, higher on field edges and higher on west than on east sides of affected fields (Collins, 1950; Gowing, 1962; Newett and Rigden. 2015; Py et al., 1987). No data were found on the extent of fruit losses associated with crown injury, but Collins (1950) reported the incidence in different fields ranged from 1.0 to 22.0%.

Maturing pineapple fruits are highly susceptible to solar injury (SI; sunburn) and several methods are used to reduce or prevent such injury and associated losses (Bartholomew, 2008; Chavarria and Ramirez, 2008; Newett and Rigden, 2015; Phillips and Bell, 2008). However, no references were found on losses of fruits due to SI injury to crowns and few formal reports of research on the subject were found. No information was found on the period when crowns are most susceptible, what the lethal tissue temperature is or how to control the problem.

Crown injury surely is a greater problem for the fresh fruit industry than for the fruit processing industry because crowns are part of formal marketing and quality standards (UNECE; USDA). When the inflorescence apical tissue is destroyed by SI before crown leaf tissues are fully developed the result is fruits without crowns (Newett and Rigden, 2015). When crown apical meristem tissues are killed later during crown development one or more buds in crown leaf axils are released resulting in a variety of crown abnormalities, including fruit with multiple crowns (Figure 1; Newett and Rigden, 2015). Severe abnormalities, including multiple crowns that can't be trimmed to meet marketing standards or that result in misshapen fruits, cause fruits to be unmarketable..

The above review was developed to help understand why approximately 30 ha of Oahu, Hawaii fields of 'CO-2' (a.k.a. 73-50) pineapple had a high incidence of crown abnormalities. The blocks were forced with ethephon in late May and early to mid June and harvested in late November and early to mid December 2015. In some blocks up to 25% of the fruits had crown abnormalities so severe they were unmarketable. As a result 8,450 boxes of pineapples did not meet marketing standards (Y. Rosa, personal communication). The problem, and especially its magnitude, had not previously been encountered in fields of pineapple grown on Oahu. Fruits that do not meet marketing standards can only be used for fresh cut fruit or juiced and both products are of lower value or have a limited market, or both.

The crown apical meristem had been killed on fruits that had crown abnormalities (Figure 1B, C, D) so it was initially thought to be the result of a spray application error. However, spray errors were quickly ruled out because the size of the affected area was so large that the highly experienced spray crew would have to have made multiple application errors, which was just not possible. More significantly, no sprays containing phytotoxic compounds had been applied to the affected area and the pattern of injury was asymmetrical with the greatest incidence of multiple crowns on the west side of affected blocks.

Examination of fruits in the affected fields indicated that crown SI occurred before, during or relatively soon after anthesis (flower opening), or all of the foregoing. The most severely affected blocks were oriented northwest – southeast and the injury was most severe on the western sides of such blocks (Figure 2). Those blocks had been forced in late May through mid June, but no data had been collected on the date of emergence of the inflorescence in the leaf whorl so it was not possible to associate weather events with the incidence of SI.

There was some evidence that SI was cultivar specific. Some blocks planted to an 'MD-2' clone located at a lower elevation had been forced on June 8 and June 15, 2015 and fruits in one block were scheduled for harvest during the second week of December, 2015. Blocks in the field were oriented northeast-southwest so it was expected that abnormal crowns would be most prevalent at the ends rather than the sides of the block. None

of the fruits on the eastern end of the block had abnormal crowns and only six abnormal crowns were found on a similar number of fruits on the westerly end of the block. We concluded that the greater exposure of 'CO-2' fruits, in part because of their much longer peduncle (Figure 3), accounted for much of the difference between the two cultivars. However, the extensive foliage in the ration field of 'MD-2' plants also provided shade for fruits of that cultivar.

Presently there is no known control for crown SI that would be suitable for large farms. However, if SI incidence is high enough to justify some research, it may be that kaolin reflectants such as Surround® WP will provide some protection. It remains to be seen whether the protectant will be washed or weathered off by harvest time so the fruit will appearance will not be affected.

#### References

- Bartholomew, D.P. Control of fruit sunburn in Taiwan. Pineapple News No. 15, p. 6. <a href="http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews15.pdf">http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews15.pdf</a>.
- Bartholomew, D.P., Malezieux, E., Sanewski, G.M., and Sinclair, E., 2003. Inflorescence and fruit development and yield. p. 167-202. In: Bartholomew, D. P., Paull, R., and Rohrbach, K. G. (eds.), The Pineapple: Botany, Production and Uses. CABI Publishing, Wallingford.
- Chavarria G.A. and E.F. Ramirez. 2008. Saran shade cloth to prevent sunburn in pineapple fruit. Pineapple News 15: 21-23. <a href="http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews15.pdf">http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews15.pdf</a>.
- Collins, J.L. 1950. The nature of multiple crowns. Pineapple Research Institute of Hawaii. PRI Special Report 21.
- Gowing, D.P., 1962. Some experiments on the influence of several factors on fasciation and multiple crowns. Pineapple Research Institute of Hawaii PRI News 10:147 157.
- Newett, S. and P. Rigden. 2015. The pineapple problem solver field guide. Horticulture Innovation Australia. Queensland. Department of Agriculture and Fisheries. 324 pages.
- Phillips, N. and D. Bell. 2004. Surround WP Crop Protectant for the Reduction of Sunburn Damage in Pineapple. Pineapple News 11:8-9. <a href="http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews11.pdf">http://www.ishs-horticulture.org/workinggroups/pineapple/PineNews11.pdf</a>.
- Py, C., Lacoeuilhe, J.J., and Teisson, C., 1987. The pineapple. Cultivation and uses. Editions G.-P. Maisonneuve, Paris.
- UNECE Standard on the marketing and commercial quality control of pineapples.

  <a href="https://www.unece.org/fileadmin/DAM/trade/Publications/ECE\_TRADE\_398E\_PineappleBrochure">https://www.unece.org/fileadmin/DAM/trade/Publications/ECE\_TRADE\_398E\_PineappleBrochure</a>

  pdf
- USDA pineapple grades and standards. <a href="https://www.ams.usda.gov/grades-standards/pineapple-grades-and-standards">https://www.ams.usda.gov/grades-standards/pineapple-grades-and-standards</a>.

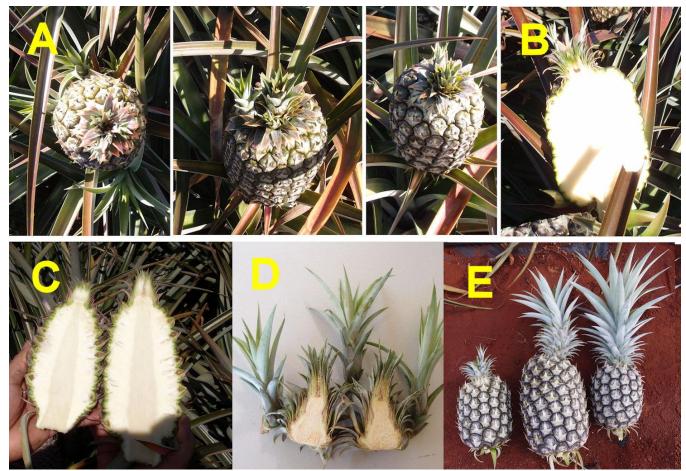


Figure 1. A. Examples of crown abnormalities on fruits of 'CO-2' after the crown stem apex was killed by heat. B. Cross section of crown on fruit near flat eye stage. C. Younger fruits with uninjured crown apex on left and heat killed apex on right. D. Triple crown formed after heat killed the apex of the crown stem. E. At the left is a fruit with a small crown after the original crown stem apex killed by heat; middle, a fruit with a single offset crown that developed after the original crown apex was killed; the fruit probably meets most market standards; at right a fruit with a normal crown.



Figure 2. On December 12, 2015 there were 8 abnormal crowns in 208 fruits (3.8%) on the east side of block B8 – 34 and 25 abnormal crowns on 225 fruits (11.1%) on the west side of the block.

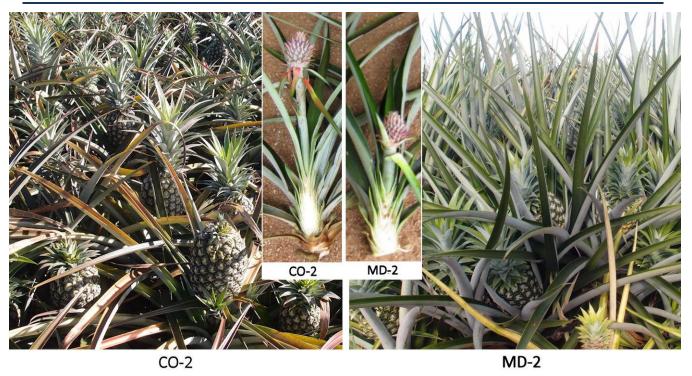


Figure 3. Fruit exposure and peduncle length of a mother plant crop of 'CO-2' and a vigorous ration crop of 'MD-2'. Much of the difference in fruit exposure is due to the much longer peduncle supporting fruits of 'CO-2'.

# **Services**

The listings below are provided as a convenience to readers and should in no way be construed as an endorsement of those providing commercial or professional services. Those offering specialized services to pineapple growers or researchers are invited to contact the editor for possible inclusion in the listings below.

## **Commercial Services**

Maintain CF 125 continues to be available for use in pineapple plant propagation anywhere in the world. Supplies can be obtained from N. Bhushan Mandava, Repar Corporation, 8070 Georgia Ave., Suite 209, Silver Spring, MD 20910. Tel: (301) 562 – 7330; Fax: (202) 223 – 0141; On the web at www.reparcorp.com; E-Mail: mandava@compuserve.com.

# **Professional Services**

Dr. Mark Paul Culik. INCAPER, Rua Alfonso Sarlo 160, CEP 29052-010, Vitoria, ES, Brazil; Tel: 27-3636-9817; Email: markculik3@yahoo.com. Experience: PhD in Plant and Soil Sciences with more than 25 years of agricultural pest management experience in crops ranging from apples to papaya and pineapple, identification of pests and beneficial arthropods ranging from mites to fruit flies, and current work on scale insects, including pineapple mealybugs. Areas of specialization: Entomology, Insect and Pest Identification, Integrated Pest Management.

Dr. Herve Fleisch. Interested in consulting on most agronomic and managerial aspects of production operations. See online profile at <a href="http://www.linkedin.com/pub/herve-fleisch/28/536/21a">http://www.linkedin.com/pub/herve-fleisch/28/536/21a</a>

Mr. Rob Moss. E-mail: <u>robmoss@bioteq-ouest.com</u>. I have 30 years experience as a tropical agronomist, have worked with pineapple since 2004 and am now helping Ghana pineapple export companies improve yields and production efficiency. I authored articles in Pineapple News No. 17, pp. 23 (Pineapple and carbon emissions); 20, pp. 57 – 65 (Greenhouse gas emissions of pineapple); and 21, pp. 40-45 (Integrated approach to disease control & soil sertility management for 'MD-2' pineapple) and am an expert on microbiological crop amendments. I am currently testing their potential to increase yields of MD-2 pineapples.

Ing. Jhonny Vasquez Jimenez, MSc. San Carlos, Costa Rica. E-mail: jvasquez@proagrocr.com, Phone: (506) 89103878, (506) 24756795. Advice on the agricultural management of pineapple crop. Analysis and improvement of pineapple crop systems for producer companies (environment and productive potential, nutrition, control pathology, crop management). For Agrochemical Companies, designing and conducting researches for new production technologies in the area of nutrition, plant pathology, weeds and other disorders.

# **Book Reviews and Web Sites**

## **Book Reviews**

No reviews were provided for this issue.

## **Web Sites of Possible Interest**

# **New References on Pineapple**

The list below includes papers related to various aspects of pineapple culture, physiology, processing, preservation or byproducts that were published or located for the period since the last issue up to about March 31, 2013. Some papers may seem relatively unrelated to pineapple but the list follows the principle of inclusion to provide the widest possible content. Often, abstracts of the papers listed below can be found on-line. I suggest searching using the paper title. Of course all abstracts of papers published in Acta Horticulturae are available from <a href="maintied">info@ishs.org</a>. For a larger view, adjust the magnification in Adobe Reader.

- Achigan-Dako, E.G., Adjé, C.A., Danikou, S.N., Hotegni, N.V.F., Agbangla, C., and Ahanchédé, A., 2014. Drivers of conservation and utilization of pineapple genetic resources in Benin. *SpringerPlus*:1-11.
- Adegbite, O. and Adeoye, I.B., 2015. Technical efficiency of pineapple production in Osun state, Nigeria. AGRIS On-line Papers in Economics and Informatics:3-12.
- Adepoju, A.O., Owoeye, I.T., and Adeoye, I.B., 2015. Determinants of market participation among pineapple farmers in Aiyedaade Local Government Area, Osun State, Nigeria. *International Journal of Fruit Science* 15:392-404.
- Adorno, W.T., Almeida, L.d.J., Martins, G.A.d.S., Silva, W.G.d., and Oliveira, P.A., 2015. Mathematics modeling applied to the surface area of pineapple pearl. *Engevista* 17:35-43.
- Afolabi, O., Oloyede, A., Abibu, W., and Adeyanju, A., 2015. Microbial safety of polyethylene packaged sliced fruits sold in Abeokuta, South-West Nigeria. *Journal of Natural Sciences Research* 5:16-21.
- Agbangba, C.E., Dagbenonbakin, G.D., Djogbenou, C.P., Houssou, P., Assea, E.D., Sossa, E.L., Kotomalè, U.A., Ahotonou, P., Ndiaga, C., and Akpo, L.E., 2015. Influence of mineral fertilizers on the physicochemical quality and organoleptic properties of pineapple cv. Smooth Cayenne in Benin. *International Journal of Biological and Chemical Sciences* 9:1277-1288.
- Akinola, O.O., 2014. Review of the role of plant in carbondioxide sequestration globally using chlorophy II or leaf index. *Journal of Environment and Earth Science* 4:22-30.
- Almeida, J.C.S., Figueiredo, D.M.d., Boari, C.A., Paixão, M.L., Sena, J.A.B., Barbosa, J.L., Ortêncio, M.O., and Moreira, K.F., 2015. Performance, body measurements, carcass and cut yields, and meat quality in lambs fed residues from processing agroindustry of fruits. *Semina: Ciências Agrárias (Londrina)* 36:541-555.
- Alphonce, R., Temu, A., and Almli, V.L., 2015. European consumer preference for African dried fruits. *British Food Journal* 117:1886-1902.
- Alvarez, R.A., Martin, R.R., and Quito-Avila, D.F., 2015. First report of Pineapple mealybug wilt associated virus-1 in Ecuador. *New Disease Reports* 31:15.
- Amaral, U.d., Maia, V.M., Pegoraro, R.F., Kondo, M.K., and Maia, L.C.B., 2015. Dry matter, carbon and nitrogen content in irrigated culture of 'Pérola' pineapple. *Interciencia* 40:639-643.
- Amenaghawon, N.A., Oronsaye, J.E., and Ogbeide, S.E., 2014. Statistical optimisation of fermentation conditions for citric acid production from pineapple peels. *Nigerian Journal of Technological Research* 9:20-26.
- Animesh, D. and Chaudhuri, P.S., 2014. Earthworm community structure of pineapple (*Ananas comosus* ) plantations under monoculture and mixed culture in West Tripura, India. *Tropical Ecology* 55:1-17.
- Anonymous, 2014. Plant variety descriptions submitted for registration of plant breeders' rights in Australia up to 30 April 2014 (includes *Ananas comosus* cv. Aus-Festival). *Plant Varieties Journal* 27:376 pp.
- Archibong, E.J., Ezemba, C.C., Chukwujama, I.C., and Archibong, U.E., 2015. Production of wine from mixed fruits: Pineapple (*Ananas comosus*) and Orange (Citrus sinensis) using yeast isolated from palm wine. *World Journal of Pharmacy and Pharmaceutical Sciences (WJPPS)* 4:126-136.
- Arinloye, D.D.A.A., Pascucci, S., Linnemann, A.R., Coulibaly, O.N., Hagelaar, G., and Omta, O.S.W.F., 2015. Marketing channel selection by smallholder farmers. *Journal of Food Products Marketing* 21:337-357.
- Arrieche, N., Paz, R., and Nogales, M., 2015. Diversity of the group Parasítica (Hymenoptera: Apócrita) associated with pineapple crop in Lara State, Venezuela. *Bioagro* 27:51-56.
- Asante, M.K. and Kuwornu, J.K.M., 2014. A comparative analysis of the profitability of pineapple-mango blend and pineapple fruit juice processing in Ghana. *APSTRACT: Applied Studies in Agribusiness and Commerce*:33-42.
- Astiani, D., Mujiman, Hatta, M., Hanisah, and Fifian, F., 2015. Soil CO2 respiration along annual crops or land-cover type gradients on West Kalimantan degraded peatland forest. *Procedia Environmental Sciences* 28:132-141.
- Aular, J., Casares, M., and Natale, W., 2014. Mineral nutrition and fruit quality of pineapple and passion fruit. *Revista Brasileira de Fruticultura* 36:1046-1054.
- Bahiense, D.V. and Souza, P.M.d., 2015. Incentives to the development of fruit crops in the Rio de Janeiro State. *Pesquisa Agropecuária Tropical* 45:113-121.
- Bahiense, D.V., Souza, P.M.d., and Ponciano, N.J., 2015. Incentives to the fruit production and the changes in the agricultural of the northern region of Rio de Janeiro. *Revista Brasileira de Fruticultura* 37:387-395.
- Baite, D.J., Chanu, T.M., Singh, M.K., Ram, D., Sangeeta, A., and Bishwapati, D., 2014. Factors related with adoption of pineapple cultivation practices by the tribal farmers of Manipur. *International Journal of Tropical Agriculture* 32:847-849.
- Baruwa, O.I., 2013. Profitability and constarints of pineapple production in Osun State, Nigeria. *Journal of Horticultural Research* 21:59-64.
- Bataglion, G.A., Silva, F.M.A.d., Eberlin, M.N., and Koolen, H.H.F., 2015. Determination of the phenolic composition from Brazilian tropical fruits by UHPLC-MS/MS. *Food Chemistry* 180:280-287.

- Batamoussi, M.H., B.S.F., X., Tokore, J.S.B.O.M., Babayai O, L.L., and Tovihoudji, P.G., 2015. Contribution to the improvement of the farming pratices of production of pineapple in Benin: case of the municipality of Aada in the department of the Atlantic. *European Scientific Journal* 11:315-326.
- Bevilacqua, A., Campaniello, D., Sinigaglia, M., and Corbo, M.R., 2015. Combination of ultrasound and antimicrobial compounds towards Pichia spp. and Wickerhamomyces anomalus in pineapple juice. *LWT Food Science and Technology* 64:616-622.
- Bhoite, A.A., Unde, K.B., Walke, A.K., Yeola, P.R., and Yewale, N.S., 2015. Studies on dehydration of pineapple using different sugar syrup treatments. *Food Science Research Journal* 6:63-66.
- Bhol, N., Sarangi, S.K., and Behera, U.K., 2015. Coconut (Cocos nucifera)-based farming system: a viable land use option for small and marginal farmers in coastal Odisha. *Indian Journal of Agricultural Sciences* 85:1488-1497.
- Bitzer, V. and Bijman, J., 2015. From innovation to co-innovation? An exploration of African agrifood chains. *British Food Journal* 117:2182-2199.
- Buah, J.N., Asare, P.A., and Arthur Junior, R., 2015. In vitro growth and multiplication of pineapple under different duration of sterilization and different concentrations of benzylaminopurine and sucrose. *Biotechnology* 14:35-40.
- Buitrago, B., Jaramillo, F., and Gómez, M., 2015. Some properties of natural fibers (sisal, pineapple, and banana) in comparison to manmade technical fibers (aramide, glass, carbon). *Journal of Natural Fibers* 12:357-367.
- Caetano, L.C.S., Ventura, J.A., and Balbino, J.M.d.S., 2015. Behavior of pineapple genotypes fusariose resistant compared to susceptible commercial cultivars. Revista Brasileira de Fruticultura 37:404-409.
- Chacón, P., Lorenz, K., Lal, R., Calhoun, F.G., and Fausey, N.R., 2015. Association of soil organic carbon with physically separated soil fractions in different land uses of Costa Rica. *Acta Agriculturæ Scandinavica, Section B Soil & Plant Science* 65:448-459.
- Chaurasiya, R.S., Sakhare, P.Z., Bhaskar, N., and Hebbar, H.U., 2015. Efficacy of reverse micellar extracted fruit bromelain in meat tenderization. *Journal of Food Science and Technology (Mysore)* 52:3870-3880.
- Cheah, P.M., Husni, M.H.A., Samsuri, A.W., and Chuah, A.L., 2013. Short-term field decomposition of pineapple stump biochar in tropical peat soil. *Malaysian Journal of Soil Science* 17:85-97.
- Chopart, J.L., Debaut-Henoque, L., Marie-Alphonsine, P.A., Asensio, R., and Soler, A., 2015. Estimating root length density of pineapple (*Ananas comosus* (L.) Merr.) from root counts on soil profiles in Martinique (French West Indies). *Fruits (Paris)* 70:143-151.
- Coppens d'Eeckenbrugge, G. and Govaerts, R., 2015. Synonymies in Ananas (Bromeliaceae). Phytotaxa 239: 273-279.
- Conesa, C., García-Breijo, E., Loeff, E., Seguí, L., Fito, P., and Laguarda-Miró, N., 2015. An Electrochemical Impedance Spectroscopy-based technique to identify and quantify fermentable sugars in pineapple waste valorization for bioethanol production. *Sensors* 15:22941-22955.
- Cruz, H., Herrera, D., and Murillo, A., 2013. Growth and absorption of nutrients in pineapple (*Ananas comosus*, var. MD-2) in the humid tropics of Costa Rica. *Tierra Tropical: Sostenibilidad, Ambiente y Sociedad* 9:11-18.
- Cruz, L.I.B., Cruz, M.d.C.M., Castro, G.D.M.d., Fagundes, M.C.P., and Santos, J.B.d., 2015. Growth and nutrition of 'Imperial' pineapple nursery plants associated with the fungus Piriformospora indica and herbicide application. *Semina: Ciências Agrárias (Londrina)* 36:2407-2422.
- Cruz, L.I.B., Cruz, M.d.C.M., Ferreira, E.A., Castro, G.D.M.d., and Almeida, M.d.O., 2014. Quantum efficiency of photosystem II of 'Imperial' pineapple nursery in response to association with the Piriformospora indica and herbicide. *Revista Brasileira de Fruticultura* 36:794-804.
- Dai, Z., Li, R., Muhammad, N., Brookes, P.C., Wang, H., Liu, X., and Xu, J., 2014. Principle component and hierarchical cluster analysis of soil properties following biochar incorporation. *Soil Science Society of America Journal* 78:205-213.
- Dang, Y. and Zhu, C., 2015. Genomic study of the absorption mechanism of p-coumaric acid and caffeic acid of extract of *Ananas comosus* L. leaves. *Journal of Food Science* 80:C504-C509.
- Dantas, A.L., Silva, S.d.M., Dantas, R.L., Pereira, W.E., Lima, R.P., Mendonça, R.M.N., and Santos, D., 2015. Influence of combined sources of nitrogen fertilization on quality of cv. Vitória pineapple. *African Journal of Agricultural Research* 10:3814-3824.
- Daramola, F., Afolami, S., Enikuomehin, O., Omonhinmin, C., and Adebayo, A., 2015. Nematicidal effects of carbofuran and GC-MS analysis of its residue in pineapple fruits. *International Journal of Agriculture and Biology* 17:357-362.
- Debasis, B., Munsi, P.S., and Ray, D.P., 2014. Studies on the performance of intercrops on the nutrient uptake and yield attributes of mango cv. AMRAPALI. *Advance Research Journal of Crop Improvement* 5:69-73.
- Dejarme-Calalang, G.M., Bock, L., and Colinet, G., 2015. Crop production of Northern Mindanao, Philippines: its contribution to the regional economy and food security. *Tropicultura* 33:77-90.
- Deng, M. and Luo, Y., 2014. Evaluation of heavy metals contamination on representative fruits in Nanning. *Meteorological and Environmental Research* 5:22-23.
- Ding, P. and Syazwani, S., 2015. Maturity stages affect antioxidant activity of 'MD2' pineapple (Ananas comosus L.).223-226.
- Donkor, A., Osei-Fosu, P., Nyarko, S., Kingsford-Adaboh, R., Brajesh, D., and Asante, I., 2015. Validation of QuEChERS method for the determination of 36 pesticide residues in fruits and vegetables from Ghana, using gas chromatography with electron capture and pulsed flame photometric detectors. *Journal of Environmental Science and Health. Part B, Pesticides, Food Contaminants, and Agricultural Wastes* 50:560-570.
- Effiom, D.O., 2015. Correlation studies of mineral nutrients' concentrations in soils and pineapple (*Ananas comosus*) plants in the southern agricultural zone of Cross River State. *Global Journal of Agricultural Sciences* 14:51-59.
- Espinosa-Rodríguez, C.J., Nieto-Angel, D., León-García de Alba, C.d., Villegas-Monter, Á., Aguilar-Pérez, L.A., and Ayala-Escobar, V., 2015. Etiology of the heart rot of pineapple (*Ananas comosus L. Merril*) MD2 cultivar in Isla, Veracruz, México. *Revista Mexicana de Fitopatología* 33:104-115.
- Ezeaku, P.I., 2015. Evaluation of agro-ecological approach to soil quality assessment for sustainable land use and management systems. Scientific Research and Essays 10:501-512.
- Falcão Filho, R.d.S., Gusmão, R.P.d., Silva, W.P.d., Gomes, J.P., Carvalho Filho, E.V., and El-Aouar, A.A., 2015. Osmotic dehydration of pineapple stems in hypertonic sucrose solutions. *Agricultural Sciences* 6:916-924.

- Fanou, A., Baïmey, H., Zandjanakou-Tachin, M., and Lawouin, L., 2014. Effectiveness of botanical extracts and Cydim Super in the fight against mealybug (Dysmicoccus brevipes) associated with wilt disease in pineapple. *International Journal of Biological and Chemical Sciences* 8:2007-2014.
- Feitosa, H.O., Amorim, A.V., Lacerda, C.F., and Silva, F.B., 2011. Crescimento e extração de micronutrientes em abacaxizeiro 'vitória'. . *Revista Brasileira de Fruticultura* 33:706-712.
- Feng, K., Hu, W., Jiang, A., Sa Ren Gao, W., and Xu, Y., 2015. Effect of chitosan edible coating incorporated with origanum essential oil on preservation of fresh-cut pineapple. *Journal of Food Safety and Quality* 6:2475-2481.
- Feng, K., Hu, W., Jiang, A., Xu, Y., Sarengaowa, Li, X., and Bai, X., 2015. Growth potential of Listeria monocytogenes and Staphylococcus aureus on fresh-cut tropical fruits. *Journal of Food Science* 80:M2548-M2554.
- Ferreira, T.d.F., Souza, R.M., Ferreira, K.D.d.S., and Idalino, W.S.S., 2015. Interaction of Rotylenchulus reniformis and Meloidogyne javanica with mealybug wilt of pineapple, in microplots. *European Journal of Plant Pathology* 141:761-768.
- Ferreira, T.d.F., Souza, R.M., Idalino, W.S.S., Ferreira, K.D.d.S., and Brioso, P.S.T., 2014. Interaction of Pratylenchus brachyurus and Helicotylenchus sp. with mealybug wilt of pineapple in microplots. *Nematropica* 44:181-189.
- Fontes, C.P.M.L., Silva, J.L.A.d., Rabelo, M.C., and Rodrigues, S., 2015. Development of low caloric prebiotic fruit juices by dextransucrase acceptor reaction. *Journal of Food Science and Technology (Mysore)* 52:7272-7280.
- Fournier, P., Benneveau, A., Hardy, C., Chillet, M., and Léchaudel, M., 2015. A predictive model based on a pluviothermic index for leathery pocket and fruitlet core rot of pineapple cv. 'Queen'. *European Journal of Plant Pathology* 142:449-460.
- Gambin, J.S. and Herrera, D., 2012. Nutrient absorption curves for pineapple (*Ananas comosus* var. MD-2). *Tierra Tropical: Sostenibilidad, Ambiente y Sociedad* 8:169-178.
- Gambley, C. and Thomas, J., 2015. Mealybug wilt disease, p. 228-236, CABI Plant Protection Series, No.3. CABI, Wallingford.
- Garcia, W.M., Krause, W., Araújo, D.V.d., Silva, C.A., and Miranda, A.F.d., 2015. Behavior in vitro of the Fusarium guttiform and evaluation of the inoculation methods in the leaves of pineapple. *Revista Caatinga* 28:263-268.
- García-Caicedo, M.M., Giraldo-Vanegas, H., and Ochoa, Á., 2012. Life cycle of Metamasius dimidiatipennis Champeon (Coleoptera: Curculionidae) under laboratory conditions. *Agronomía Tropical (Maracay)* 62:69-75.
- Gowda, N.K.S., Vallesha, N.C., Awachat, V.B., Samireddypalli, A., Pal, D.T., and Prasad, C.S., 2015. Study on evaluation of silage from pineapple (*Ananas comosus*) fruit residue as livestock feed. *Tropical Animal Health and Production* 47:557-561.
- Green, J. and Nelson, S. 2016. Heart and Root Rots of Pineapple. College of Tropical Agriculture and Human Resources (CTAHR), Cooperative Extension Service/CTAHR, University of Hawai'i at Mānoa, Honolulu, Hawai'i 96822.
- Gu, H., Zhan, R.L., Zhang, L.B., Gong, D.Q., and Jia, Z.W., 2015. First report of Fusarium ananatum causing pineapple fruitlet core rot in China. *Plant Disease* 99:1653.
- Guácimo, L.M., 2012. Curvas de absorción de nutrientes en el cultivo de piña. *Tierra Tropical: Sostenibilidad, Ambiente y Sociedad* 36:169-178.
- Haque, M.A., Afrin, S., Shibly, A.Z., Zohora, F.T., and Sultana, M.M., 2015. Effect of pineapple pulp on sensory quality and shelf-life of pineapple cake. *Journal of Environmental Science and Natural Resources* 8:7-11.
- He, Y.B., Zhan, R.L., Sun, G.M., Wu, J.B., and Zhao, Y.L., 2015. Phylogeography of pink pineapple mealybugs, Dysmicoccus brevipes (Cockerell) reveals the history of pineapple introduction and cultivation in China. *Genetics and Molecular Research* 14:9890-9897.
- Hegde, G.M., 2014. Integrated management of heart rot of pineapple. The Bioscan 9:1229-1232.
- Hernández, G.A., Tello, N., and Herrera, D., 2012. Pineapple (*Ananas comosus*, var. MD-2) growth on a farm in the humid tropics of Costa Rica. *Tierra Tropical: Sostenibilidad, Ambiente y Sociedad* 8:159-167.
- Hotegni, V.N.F., 2014. Using agronomic tools to improve pineapple quality and its uniformity in Benin. Wageningen Universiteit (Wageningen University), Wageningen, Ph.D.
- Hotegni, V.N.F., Lommen, W.J.M., Agbossou, E.K., and Struik, P.C., 2014. Heterogeneity in pineapple fruit quality results from plant heterogeneity at flower induction. *Frontiers in Plant Science* 5:670.
- Hotegni, V.N.F., Lommen, W.J.M., Agbossou, E.K., and Struik, P.C., 2015. Influence of weight and type of planting material on fruit quality and its heterogeneity in pineapple [Ananas comosus (L.) Merrill]. Frontiers in Plant Science 6:798.
- Hotegni, V.N.F., Lommen, W.J.M., Agbossou, E.K., and Struik, P.C., 2015. Selective pruning in pineapple plants as means to reduce heterogeneity in fruit quality. *SpringerPlus* 4:(14 March 2015).
- Hounhouigan, M.H., 2014. Quality of pasteurised pineapple juice in the context of the Beninese marketing system. Wageningen Universiteit (Wageningen University), Wageningen.
- Hu, J., Li, X., Luo, Z., Fan, H., Zhang, Z., Liu, Z., and He, F., 2015. Detection of Pineapple mealybug wilt associated virus-3 by Real-Time Fluorescent Quantitative RT-PCR. *Journal of Fruit Science* 32:156-162.
- Hu, W., Liu, S., and Liaw, S., 2015. Long-term preconditioning of plantlets: a practical method for enhancing survival of pineapple (*Ananas comosus* (L.) Merr.) shoot tips cryopreserved using vitrification. *CryoLetters* 36:226-236.
- Ibeawuchi, I.I., Okoli, N.A., Alagba, R.A., Ofor, M.O., Emma-Okafor, L.C., Peter-Onoh, C.A., and Obiefuna, J.C., 2015. Fruit and vegetable crop production in Nigeria: the gains, challenges and the way forward. *Journal of Biology, Agriculture and Healthcare* 5:194-208.
- Ibrahim, N.F., Mohd, M.H., Nor, N.M.I.M., and Zakaria, L., 2015. First report of Fusarium oxysporum and F. solani associated with pineapple rot in Peninsular Malaysia. *Plant Disease* 99:1650.
- Islam, M.A., Ahmad, I., Ahmed, S., and Sarker, A., 2014. Biochemical composition and shelf life study of mixed fruit juice from orange & pineapple. *Journal of Environmental Science and Natural Resources* 7:227-232.
- Jayaprakashvel, M., Akila, S., Venkatramani, M., Vinothini, S., Bhagat, S.J., and Hussain, A.J., 2014. Production of bioethanol from papaya and pineapple wastes using marine associated microorganisms. *Biosciences, Biotechnology Research Asia* 11:193-199.
- Jayasinghe, A.P., Weerahewa, H.L.D., and Dassanayake, E.M., 2012. Optimization of the conditions for acclimatization of micropropagated pineapple propagales. 306-308.

- Kalibwani, R., Kamugisha, R., Twebaze, J., Sabiiti, M., Kugonza, I., Tenywa, M., and Nyamwaro, S.O. 2015. Multi-Stakeholder Partnerships in Organic Value Chain Development: A Case of Ntungamo Innovation Platform in Western Uganda, Lagos, Nigeria. 3rd African Organic Conference).
- Kamalakar, D., Rao, K.S., and Rao, L.N., 2015. Studies on drying characteristics of pineapple. World Journal of Pharmaceutical Research 4:428-442.
- Khayankarn, S., Jarintorn, S., Srijumpa, N., Uthaibutra, J., and Whangchai, K., 2014. Control of Fusarium sp. on pineapple by megasonic cleaning with electrolysed oxidising water. *Maejo International Journal of Science and Technology* 8:288-296.
- Krishnan, V.A. and Gokulakrishnan, M., 2015. Extraction, purification of bromelain from pineapple and determination of its effect on bacteria causing periodontitis. *International Journal of Pharmaceutical Sciences and Research (IJPSR)* 6:5284-5294.
- Lalthanzara, H. and Ramanujam, S.N., 2014. Earthworm cast production and physico-chemical properties in two agroforestry systems of Mizoram (India). *Tropical Ecology* 55:75-84.
- Leonardo, F.d.A.P., Pereira, W.E., Silva, S.d.M., Araújo, R.d.C., and Mendonça, R.M.N., 2014. Optimum size of the experimental plot of pineapple 'Victory'. *Revista Brasileira de Fruticultura* 36:909-916.
- Leone, G.F., Almeida, C.V.d., Abreu-Tarazi, M.F.d., Batagin-Piotto, K.D., Artioli-Coelho, F.A., and Almeida, M.d., 2016. Antibiotic therapy in pineapple (*Ananas comosus*) microplants. *Ciência Rural* 46:89-94.
- Li, J., Wang, Y., Yan, S., Li, X., and Pan, S., 2016. Molecularly imprinted calixarene fiber for solid-phase microextraction of four organophosphorous pesticides in fruits. *Food Chemistry* 192:260-267.
- Liang, J., Xu, J., Pan, J., Ge, M., and Zong, K., 2015. Identification of the main allergenic proteins in high hydrostatic pressure pineapple juice and assessing the influence of pressure on their allergenicity. *International Journal of Food Properties* 18:2134-2144.
- Lima, R.M.B., Sousa, W.H.d., Medeiros, A.N.d., Cezar, M.F., Cartaxo, F.Q., Gonzaga Neto, S., Costa, R.G., and Medeiros, G.R.d., 2015. Characteristics of the carcass of goats of different genotypes fed pineapple (*Ananas comosus* L.) stubble hay. *Revista Brasileira de Zootecnia* 44:44-51.
- Lin, M., Chen, M., Lin, T., Kuan, C., Lee, C., and Yang, W., 2015. Prevention of natural flowering in pineapple (*Ananas comosus*) by shading and urea application. *Horticulture, Environment and Biotechnology* 56:9-16.
- Lins, A.D.F., Lisbôa, C.G.C.d., Moraes, M.S.d., Sampaio, A.C.F., and Quirino, D.J.G., 2015. Microbiological analysis minimally processed fruits Food and Nutrition Unit. Revista Verde de Agroecologia e Desenvolvimento Sustentável 10:Ciencia de Alimentos 22-25.
- Lorenzo, J.C., Yabor, L., Medina, N., Quintana, N., and Wells, V., 2015. Coefficient of variation can identify the most important effects of experimental treatments. *Notulae Botanicae, Horti Agrobotanici, Cluj-Napoca* 43:287-291.
- Ma, J., Kanakala, S., He, Y., Zhang, J., and Zhong, X., 2015. Transcriptome sequence analysis of an ornamental plant, *Ananas comosus* var. bracteatus, revealed the potential unigenes involved in terpenoid and phenylpropanoid biosynthesis. *PLoS ONE* 10:e0119153.
- Machado, A.C.Z., Siqueira, K.M.S., Ferraz, L.C.C.B., Inomoto, M.M., Bessi, R., Harakava, R., and Oliveira, C.M.G., 2015. Characterization of Brazilian populations of Pratylenchus brachyurus using morphological and molecular analyses. *Tropical Plant Pathology* 40:102-110.
- Madihah Binti, A.G., Rozita, N., Javad Hamzehalipour, A., Fadzilah Adibah, A.M., Mohsen, M., Neda, A., Siavash Hosseinpour, C., and Mirdawati, M., 2015. In vitro antiproliferative activity of fresh pineapple juices on ovarian and colon cancer cell lines. *International Journal of Peptide Research and Therapeutics* 21:353-364.
- Mahadwar, G., Chauhan, K.R., Bhagavathy, G.V., Murphy, C., Smith, A.D., and Bhagwat, A.A., 2015. Swarm motility of Salmonella enterica serovar Typhimurium is inhibited by compounds from fruit peel extracts. *Letters in Applied Microbiology* 60:334-340.
- Manzocco, L., Plazzotta, S., Maifreni, M., Calligaris, S., Anese, M., and Nicoli, M.C., 2016. Impact of UV-C light on storage quality of fresh-cut pineapple in two different packages. *LWT Food Science and Technology* 65:1138-1143.
- Matos, R.M.d., Medeiros, R.M.d., Francisco, P.R.M., Silva, P.F.d., and Santos, D., 2015. Characterization and climatic aptitude of crops for municipality of the Alhambra-PB, Brazil. *Revista Brasileira de Agricultura Irrigada* 9:183-192.
- Mendes, P.d.S., Araújo, W.F., Antunes, F., Chagas, E.A., and Couceiro, M.A., 2015. In vitro cultivation of pineapple seedlings using filters, artificial ventilation and sucrose. *Agro@mbiente On-line* 9:202-207.
- Ming, R.a.m.o., 2015. The pineapple genome and the evolution of CAM photosynthesis. *Nature Genetics* 47:1435-1442.
- Minyahil, K., 2015. Survey of insects and diseases of fruit crops in SNNPR: in the case of Kafa Zone. *Journal of Biology, Agriculture and Healthcare* 5:73-81.
- Mojica, M.I. and Leblanc, H., 2013. Evaluation of humic substances and compost prepared using stubble from pineapple plantations for the reduction of nematode Radopholus simili populations in a banana crop. *Tierra Tropical: Sostenibilidad, Ambiente y Sociedad* 9:39-47.
- Mongi, R.J., Ndabikunze, B.K., Wicklund, T., Chove, L.M., and Chove, B.E., 2015. Effect of solar drying methods on total phenolic contents and antioxidant activity of commonly consumed fruits and vegetable (mango, banana, pineapple and tomato) in Tanzania. *African Journal of Food Science* 9:291-300.
- Nashima, K., Terakami, S., Nishitani, C., Kunihisa, M., Shoda, M., Takeuchi, M., Urasaki, N., Tarora, K., Yamamoto, T., and Katayama, H., 2015. Complete chloroplast genome sequence of pineapple (*Ananas comosus* ). *Tree Genetics and Genomes* 11:60.
- Nogueira, S.R., Lima, F.S.O., Rocha, E.M., and Araújo, D.H.M., 2014. Fungicides in fusariosis pineapple control in the state of Tocantins, Brazil. *Revista de Ciências Agrárias (Portugal)* 37:447-455.
- Noor Hydayaty, M.Y., Ong, W., Raimi Mohamed, R., Mariam Abd, L., and Kumar, S.V., 2015. Discovery of precursor and mature microRNAs and their putative gene targets using high-throughput sequencing in pineapple (*Ananas comosus* var. *comosus*). *Gene* 571:71-80
- Nor, M.Z.M., Ramchandran, L., Duke, M., and Vasiljevic, T., 2015. Characteristic properties of crude pineapple waste extract for bromelain purification by membrane processing. *Journal of Food Science and Technology (Mysore)* 52:7103-7112.
- Nyantika, M.M. and Aming'a, N.N., 2015. Effects of education attainment and previous training on farm practices among pineapple farmers in three barangays in Philippines. *Asian Journal of Agricultural Extension, Economics and Sociology* 5:192-201.

- Nzabuheraheza, F.D. and Nyiramugwera, A.N., 2014. Golden wine produced from mixed juices of passion fruit (Passiflora edulis), mango (Mangifera indica) and pineapple (*Ananas comosus* ). *African Journal of Food, Agriculture, Nutrition and Development* 14:9104-9116.
- Odebisi-Omokanye, M.B., Oke, M.A., El-Imam, A.M.A., Ajijolakewu, A.K., and Salaudeen, B.I., 2015. Microbiological quality and safety of pre-cut fruit retailed in Ilorin, Kwara state, Nigeria. *Fountain Journal of Natural and Applied Sciences* 4:19-26.
- Oduntan, O.B., Bamgboye, A.I., and Anisur, R., 2015. Optimization of extrusion point pressure of pineapple pomace based mash. *Agricultural Engineering International: CIGR Journal* 17:151-159.
- Olanipekun, B.F., Tunde-Akintunde, T.Y., Oyelade, O.J., Adebisi, M.G., and Adenaya, T.A., 2015. Mathematical modeling of thin-layer pineapple drying. *Journal of Food Processing and Preservation* 39:1431-1441.
- Oliveira, A.M.G., Pereira, M.E.C., Natale, W., Nunes, W.S., and Ledo, C.A.d.S., 2015. Quality of pineapple 'BRS Imperial' as a function of N-K doses. *Revista Brasileira de Fruticultura* 37:497-506.
- Oliveira, É.R., Deus, K.O.d., and Caliari, M., 2015. Production, characterization and acceptability of different alcohol-based pineapple liqueurs. *Revista Verde de Agroecologia e Desenvolvimento Sustentável* 10:Artigos 108-114.
- Oliveira, R.d.S., Pereira, M.R., Carvalho, V.S., Lucas, E.d.F., and Gravina, G.d.A., 2015. Starch and sodium hipochlorite on in vitro rooting of pineapple 'Gold' and its effects on acclimatization. *Revista Brasileira de Fruticultura* 37:273-280.
- Ostrowski, A.P., Bracht, L., Broetto-Biazon, A.C., Musial, D.C., and Ostrowski, M., 2015. Comparative study of hypocholesterolemic potential of *Ananas comosus* peels and Passiflora edulis mesocarp in rats and mice. *REBRAPA Revista Brasileira de Pesquisa em Alimentos* 6:64-69.
- Owureku-Asare, M., Agyei-Amponsah, J., Agbemavor, S.W.K., Apatey, J., Sarfo, A.K., Okyere, A.A., Twum, L.A., and Dodobi, M.T., 2015. Effect of organic fertilizers on physical and chemical quality of sugar loaf pineapple (*Ananas comosus L.*) grown in two ecological sites in Ghana. *African Journal of Food, Agriculture, Nutrition and Development* 15:9982-9995.
- Pan, Y., Zhu, J., and Li, S., 2015. Effects of pure oxygen and reduced oxygen modified atmosphere packaging on the quality and microbial characteristics of fresh-cut pineapple. *Fruits (Paris)* 70:101-108.
- Parente, G.D.L., Almeida, M.M.d., Silva, J.L.d., Silva, C.G.d., and Alves, M.F., 2014. Kinetic production of alcoholic unfermented pineapple 'Pearl' drink and characterization. *Revista Verde de Agroecologia e Desenvolvimento Sustentável* 9:Artigos 230-247.
- Parra-Matadamas, A., Mayorga-Reyes, L., and Pérez-Chabela, M.L., 2015. In vitro fermentation of agroindustrial by-products: grapefruit albedo and peel, cactus pear peel and pineapple peel by lactic acid bacteria. *International Food Research Journal* 22:859-865.
- Patil, B.S., Jayaprakasha, G.K., Roa, C.O., and Mahattanatawee, K., 2014. *Tropical and subtropical fruits: flavors, color, and health benefits.* Oxford University Press, New York.
- Patil, N.A., Yeldhalli, R.A., Patil, B.O., and Tirlapur, L.N., 2015. Impact of climate change on major fruits in India. *Asian Journal of Environmental Science* 10:34-38.
- Paull, R.E. and Chen, N.J., 2015. Pineapple translucency and chilling injury in new low-acid hybrids.61-66.
- Pellicano, R., Ribaldone, D.G., Saracco, G.M., Leone, N., Angelis, C.d., Arrigoni, A., Morello, E., Sapone, N., Cisarò, F., and Astegiano, M., 2014. Benefit of supplements in functional dyspepsia after treatment of Helicobacter pylori. *Minerva Gastroenterologica e Dietologica* 60:263-268.
- Pereira, E.M., Leite Filho, M.T., Santos, Y.M.G.d., Pereira, B.B.M., and Maracajá, P.B., 2015. Preparation and quality jelly and pineapple compote "pearl". Revista Verde de Agroecologia e Desenvolvimento Sustentável 10:Artigos 149-153.
- Pereira, M.R., Carvalho, V.S., Lucas, E.d.F., and Gravina, G.d.A., 2015. Corn starch and sodium hypochlorite on in vitro rooting of pineapple 'Vitória' and their effect on acclimatization. *Revista Brasileira de Fruticultura* 37:528-533.
- Perez C, A., Leonardo, C.A., and Romero G, J., 2014. Presence of the fungus Thielaviopsis paradoxa causing black rot of pineapple gold honey in the Department of Sucre. *Revista Colombiana de Ciencia Animal* 6:342-345.
- Pino, G.G.d., Díaz, F.V., Valin Rivera, J.L., Kieling, A.C., and Torres, A.R., 2015. Evaluation of composite materials with fiber of curauá pineapple (Ananas Erectifolius). *Review of Research Journal* 4:ROR-1355.
- Piyush, P. and Jivani, N.P., 2015. Effects of hydroalcoholic extract of Parkinsonia aculeate L. seeds and *Ananas comosus* fruits on rotenone induced Parkinson's disease in rats. *International Journal of Research and Development in Pharmacy and Life Sciences* 4:1647-1653.
- Rahimi, E. and Jeiran, M.R., 2015. Patulin and its dietary intake by fruit juice consumption in Iran. *Food Additives and Contaminants B, Surveillance* 8:40-43.
- Ramos, M.J.M. and da Rocha Pinho, L.G., 2014. Physical and Quality Characteristics of Jupi Pineapple Fruits on Macronutrient and Boron Deficiency. *Natural Resources* 5:359-366.
- Ranjitham, A.M., Ranjani, G.S., and Caroling, G., 2015. Biosynthesis, characterization, antimicrobial activity of copper nanoparticles using fresh aqueous *Ananas comosus* L. (pineapple) extract. *International Journal of PharmTech Research* 8:750-769.
- Rashid, A.Q.M.B., Meah, M.B., and Sultana, A., 2015. Seed borne fungal diseases of fruits in Mymensingh district. *Journal of Environmental Science and Natural Resources* 8:153-156.
- Redwan, R.M., Saidin, A., and Kumar, S.V., 2015. Complete chloroplast genome sequence of MD-2 pineapple and its comparative analysis among nine other plants from the subclass Commelinidae. *BMC Plant Biology* 15:(12 August 2015).
- Ribeiro, L.S., Duarte, W.F., Dias, D.R., and Schwan, R.F., 2015. Fermented sugarcane and pineapple beverage produced using Saccharomyces cerevisiae and non-Saccharomyces yeast. *Journal of the Institute of Brewing* 121:262-272.
- Russo, P., Chiara, M.L.V.d., Vernile, A., Amodio, M.L., Arena, M.P., Capozzi, V., Massa, S., and Spano, G., 2014. Fresh-cut pineapple as a new carrier of probiotic lactic acid bacteria. *BioMed Research International* 2014:Article ID 309183.
- Sabino, L.B.S., Gonzaga, M.L.C., Soares, D.J., Lima, A.C.S., Lima, J.S.S., Almeida, M.M.B., Sousa, P.H.M., and Figueiredo, R.W., 2015. Bioactive compounds, antioxidant activity, and minerals in flours prepared with tropical fruit peels. *Acta Alimentaria (Budapest)* 44:520-526.

- Sah, B.N.P., Vasiljevic, T., McKechnie, S., and Donkor, O.N., 2015. Effect of refrigerated storage on probiotic viability and the production and stability of antimutagenic and antioxidant peptides in yogurt supplemented with pineapple peel. *Journal of Dairy Science* 98:5905-5916.
- Sah, B.N.P., Vasiljevic, T., McKechnie, S., and Donkor, O.N., 2016. Physicochemical, textural and rheological properties of probiotic yogurt fortified with fibre-rich pineapple peel powder during refrigerated storage. *LWT Food Science and Technology* 65:978-986.
- Saiful Izwan, A.R., Noor Fadzliana, A.S., Nadirul Hasraf, M.N., Ida Idayu, M., and Mohd Yazid, Y., 2015. Impregnation of poly(lactic acid) on biologically pulped pineapple leaf fiber for packaging materials. *BioResources* 10:4350-4359.
- Saifullah, K., Naheed, K., Kayani, H.A., Amir Ahmed, M., and Bushra, N., 2015. Effect of grinding agents and detergents on the quality of extracted DNA from diverse plant species. *International Journal of Biology and Biotechnology* 12:39-45.
- Saikia, S., Mahnot, N.K., and Mahanta, C.L., 2015. Effect of spray drying of four fruit juices on physicochemical, phytochemical and antioxidant properties. *Journal of Food Processing and Preservation* 39:1656-1664.
- Sairi, M., Law, J.Y., and Sarmidi, M.R. 2004. Chemical composition and sensory analysis of fresh pineapple and deacidified pineapple juice using electrodialysis, Puteri Pan Pacific Hotel, Johor Bahru, Johor, Malaysia. http://eprints.utm.my/6174/.
- Saito, F., Shimizu, M., Suzuki, T., Hamada, C., Iwase, T., Okochi, N., Yamazaki, M., and Kyotani, H., 2015. Dai Nippon Printing Co., Ltd Medi.Ca CC for Enumeration of Coliform Bacteria. *Journal of AOAC International* 98:62-70.
- Salazar-López, E.I., Jiménez, M., Salazar, R., and Azuara, E., 2015. Incorporation of microcapsules in pineapple intercellular tissue using osmotic dehydration and microencapsulation method. *Food and Bioprocess Technology* 8:1699-1706.
- Salihu, S.O., Jacob, J.O., Kolo, M.T., Osundiran, B.J., and Emmanuel, J., 2014. Heavy metals in some fruits and cereals in Minna markets, Nigeria. *Pakistan Journal of Nutrition* 13:722-727.
- Salleh, J., Mohd Yusoh, M.K., and Ruznan, W.S., 2015. Tensile strength of some natural-fibre composites. *Pertanika Journal of Tropical Agricultural Science* 38:575-582.
- Sanjay, K. and Sharma, H.K., 2015. Enzymatic degumming of pineapple (*Ananas comosus*) mill juice using crude and commercial enzymes. *Journal of Food Measurement and Characterization* 9:414-425.
- Santos, C., Ventura, J.A., Costa, H., Fernandes, P.M.B., and Lima, N., 2015. MALDI-TOF MS to identify the pineapple pathogen Fusarium guttiforme and its antagonist Trichoderma asperellum on decayed pineapple. *Tropical Plant Pathology* 40:227-232.
- Santos, P.B.d., Barbosa, F.d.S., Vieira, C.F., and Carvalho, A.C.P.P., 2015. Number of explants, culture medium and photoperiod in ornamental pineapple micropropagation. *Revista Ciência Agronômica* 46:749-754.
- Scherer, R.F., Fraga, H.P.d.F., Klabunde, G.F., Silva, D.A.d., and Guerra, M.P., 2015. Global DNA methylation levels during the development of nodule cluster cultures and assessment of genetic fidelity of in vitro-regenerated pineapple plants (*Ananas comosus* var. *comosus*). *Journal of Plant Growth Regulation* 34:677-683.
- Scherer, R.F., Holderbaum, D.F., Garcia, A.C., Silva, D.A.d., Steinmacher, D.A., and Guerra, M.P., 2015. Effects of immersion system and gibberellic acid on the growth and acclimatization of micropropagated pineapple. *Crop Breeding and Applied Biotechnology* 15:66-71.
- Selani, M.M., Shirado, G.A.N., Margiotta, G.B., Saldaña, E., Spada, F.P., Piedade, S.M.S., Contreras-Castillo, C.J., and Canniatti-Brazaca, S.G., 2016. Effects of pineapple byproduct and canola oil as fat replacers on physicochemical and sensory qualities of low-fat beef burger. *Meat Science* 112:69-76.
- Shaver, I., Chain-Guadarrama, A., Cleary, K.A., Sanfiorenzo, A., Santiago-García, R.J., Finegan, B., Hormel, L., Sibelet, N., Vierling, L.A., Bosque-Pérez, N.A., DeClerck, F., Fagan, M.E., and Waits, L.P., 2015. Coupled social and ecological outcomes of agricultural intensification in Costa Rica and the future of biodiversity conservation in tropical agricultural regions. *Global Environmental Change* 32:74-86.
- Shiau, S., Wu, M., and Liu, Y., 2015. The effect of pineapple core fiber on dough rheology and the quality of mantou. *Journal of Food and Drug Analysis* 23:493-500.
- Shivabasu, K., Narayanaswamy, B., and Pampangouda, 2014. Isolation and characterization of Gluconoacetobacter sp. from different fruits. *Biochemical and Cellular Archives* 14:407-411.
- Shuvee, N., Ramesh, S.T., Gandhimathi, R., and Nidheesh, P.V., 2015. Pineapple leaf (*Ananas comosus*) powder as a biosorbent for the removal of crystal violet from aqueous solution. *Desalination and Water Treatment* 54:2041-2054.
- Sihombing, J.R., Dharma, A., Chaidir, Z., Almahdy, Fachrial, E., and Munaf, E., 2015. Phytochemical screening and antioxidant activities of 31 fruit peel extract from Sumatera, Indonesia. *Journal of Chemical and Pharmaceutical Research* 7:190-196.
- Silva, D.F.d., Pegoraro, R.F., Medeiros, A.C., Lopes, P.A.P., Cardoso, M.M., and Maia, V.M., 2015. Nitrogen and plant density in the economic evaluation and fruit quality of pineapple. *Pesquisa Agropecuária Tropical* 45:39-45.
- Silva, F.G.d., Cunha Neto, A., Rodrigues, L.J., and Figueiredo, E.E.d.S., 2015. Quality of minimally processed products marketed in Cuiabá, Mato Grosso, Brazil. *Journal of Food Research* 4:157-164.
- Silva, G.S.d., Krasucki, A.I.S., Rozário, I.L.M., and Leite, R.R., 2015. Resistance of pineapple 'Turiaçu' to Meloidogyne arenaria, M. enterolobii, M. incognita and M. javanica. *Summa Phytopathologica* 41:227-228.
- Silva, K.S., Garcia, C.C., Amado, L.R., and Mauro, M.A., 2015. Effects of edible coatings on convective drying and characteristics of the dried pineapple. *Food and Bioprocess Technology* 8:1465-1475.
- Sindhu, M., Zainul Akmar, Z., and Nur Fashya, M., 2015. Antioxidant property and chemical profile of pyroligneous acid from pineapple plant waste biomass. *Process Biochemistry* 50:1985-1992.
- Snehasis, C., Rao, P.S., and Mishra, H.N., 2015. Effect of combined high pressure-temperature treatments on color and nutritional quality attributes of pineapple (*Ananas comosus* L.) puree. *Innovative Food Science & Emerging Technologies* 28:10-21.
- Snehasis, C., Rao, P.S., and Mishra, H.N., 2015. Empirical model based on Weibull distribution describing the destruction kinetics of natural microbiota in pineapple (*Ananas comosus* L.) puree during high-pressure processing. *International Journal of Food Microbiology* 211:117-127.
- Snehasis, C., Rao, P.S., and Mishra, H.N., 2015. Response surface optimization of process parameters and fuzzy analysis of sensory data of high pressure-temperature treated pineapple puree. *Journal of Food Science* 80:E1763-E1775.

- Sossa, E.L., Amadji, G.L., Aholoukpè, N.S.H., Hounsou, B.M., Agbossou, K.E., and Hounhouigan, D.J., 2015. Change in a ferralsol physico-chemical properties under pineapple cropping system in southern of Benin. *Journal of Applied Biosciences* 91:8559-8569.
- Souto, R.F., Durigan, J.F., Santos, L.O., Souza, B.S.d., and Menegucci, J.L.P., 2014. Postharvest behavior of 'Pérola' pineapples to heat treatments cold storage. *Revista Brasileira de Fruticultura* 36:1028-1033.
- Souza, É.Á.T., Araujo, Q.R.d., Faria Filho, A.F.d., and Santana, S.O., 2012. Carbon sequestration by the main soils in Middle South, Bahia, Brazil. *Agrotrópica* 24:99-108.
- Souza, W.C.O.d., Nascimento, L.C.d., Vieira, D.L., Santos, T.S.d., and Assis Filho, F.M.d., 2015. Alternative control of Chalara paradoxa, causal agent of black rot of pineapple by plant extract of Mormodica charantia. *European Journal of Plant Pathology* 142:481-488.
- Sowjanya, K., Manjula, K., Masanam, V.S.P., and Penchalaraju, M., 2014. Physicochemical and microbial analysis of minimally processed fruits and vegetables. *Journal of Research PJTSAU* 42:40-44.
- Srivastava, P. and Saini, P., 2014. Analysis of antioxidant activity of five tropical fruits. Trends in Biosciences 7:1242-1245.
- Steingass, C.B., Carle, R., and Schmarr, H.G., 2015. Ripening-dependent metabolic changes in the volatiles of pineapple (*Ananas comosus* (L.) Merr.) fruit: I. Characterization of pineapple aroma compounds by comprehensive two-dimensional gas chromatography-mass spectrometry. *Analytical and Bioanalytical Chemistry* 407:2591-2608.
- Steingass, C.B., Glock, M.P., Schweiggert, R.M., and Carle, R., 2015. Studies into the phenolic patterns of different tissues of pineapple (*Ananas comosus* [L.] Merr.) infructescence by HPLC-DAD-ESI-MSn and GC-MS analysis. *Analytical and Bioanalytical Chemistry* 407:6463-6479.
- Steingass, C.B., Jutzi, M., Müller, J., Carle, R., and Schmarr, H.G., 2015. Ripening-dependent metabolic changes in the volatiles of pineapple (*Ananas comosus* (L.) Merr.) fruit: II. Multivariate statistical profiling of pineapple aroma compounds based on comprehensive two-dimensional gas chromatography-mass spectrometry. *Analytical and Bioanalytical Chemistry* 407:2609-2624.
- Stepien', Ł., Waśkiewicz, A., and Wilman, K., 2015. Host extract modulates metabolism and fumonisin biosynthesis by the plant-pathogenic fungus Fusarium proliferatum. *International Journal of Food Microbiology* 193:74-81.
- Subin, U., Mathew, J.J., Sajeshkumar, N.K., and Vazhacharickal, P.J., 2014. Pineapple (*Ananas comosus*) cultivation as an intercrop in rubber replant: effects on soil chemical and physical properties. *International Journal of Advanced Life Sciences (IJALS)* 7:574-579.
- Tajwinder, K., Amandeep, K., and Grewal, R.K., 2015. Kinetics studies with fruit bromelain (*Ananas comosus*) in the presence of cysteine and divalent ions. *Journal of Food Science and Technology (Mysore)* 52:5954-5960.
- Talai, S.M., Siagi, Z.O., Kimutai, S.K., Simiyu, S.S., Ngigi, W.T., and Makokha, A.B., 2014. Comparative energy generation of Irish-potato, tomato and pineapple ZN/CU vegetative batteries. Research Journal of Applied Sciences, Engineering and Technology 8:9-19.
- Taniguchi, C.A.K., Castro, A.C.R.d., Silva, T.F.d., and Café, F.B.d.S., 2015. Development of pineapple as an ornamental potted plant. *Acta Horticulture* 1087:379-384.
- Taufiq, A.M., Yusof, Y.A., Chin, N.L., Othman, S.H., Serikbaeva, A., and Aziz, M.G., 2015. Physicochemical properties of tamarind and pineapple fruit pulps and powders. *International Food Research Journal* 22:707-712.
- Taufiq, A.M., Yusof, Y.A., Chin, N.L., Othman, S.H., Serikbayeva, A., and Aziz, M.G., 2014. In-vitro dissolution of compressed tamarind and pineapple powder tablets. Agriculture and Agricultural Science Procedia 2:53-59.
- Therumthanam, A.C., Mathew, J.J., Sajeshkumar, N.K., and Vazhacharickal, P.J., 2014. Studies on pineapple (*Ananas comosus*) cultivation as an intercrop in rubber replant: effects on soil microorganisms. *Science Research Reporter* 4:119-127.
- Tossou, C.C., Capo-Chichi, D.B.E., and Yedomonhan, H., 2015. Diversity and morphological characterization varieties of pineapple (*Ananas comosus* (L.) Merrill) grown in Benin. *Journal of Applied Biosciences* 87:8113-8120.
- Troyo, R.D. and Acedo, A.L., Jr., 2015. Antimicrobial potential of lactic acid bacteria on freshcut pineapple (Ananas comosus ).413-418.
- Troyo, R.D. and Acedo, A.L., Jr., 2015. Effects of ascorbic and lactic acids on quality of freshcut pineapple (*Ananas comosus* ).425-429.
- Trujillo, J.C. and Iglesias, W.J., 2013. Measurement of the technical efficiency of small pineapple farmers in Santander, Colombia: a stochastic frontier approach. *Revista de Economia e Sociologia Rural* 51:S049-S062.
- Urasaki, N., Goeku, S., Kaneshima, R., Takamine, T., Tarora, K., Takeuchi, M., Moromizato, C., Yonamine, K., Hosaka, F., Terakami, S., Matsumura, H., Yamamoto, T., and Shoda, M., 2015. Leaf margin phenotype-specific restriction-site-associated DNA-derived markers for pineapple (*Ananas comosus* L.). *Breeding Science* 65:276-284.
- Usman, M., Elfaki, F.A.M., Wamiliana, Fauzan, and Daoud, J.I., 2015. Statistical model and prediction of pineapple plant weight. *Science International (Lahore)* 27:937-943.
- Uthairatanakij, A., Aiamla-or, S., and Jitareerat, P., 2015. Preharvest calcium effects on internal breakdown and quality of 'Pattavia' pineapple during low temperature storage. *Acta Horticulturae* 1088:443-448.
- Vilela, G.B., Pegoraro, R.F., and Maia, V.M., 2015. Predicting the production of 'Vitória' pineapple from phytotechnical and nutritional characteristics. *Revista Ciência Agronômica* 46:724-732.
- Virendra, S., Jadhav, S.B., and Singhal, R.S., 2015. Interaction of polyphenol oxidase of Solanum tuberosum with β-cyclodextrin: process details and applications. *International Journal of Biological Macromolecules* 80:469-474.
- Waal, J.W.H.v.d., 2015. Grapefruit seed extracts as organic post-harvest agents: precious lessons on efficacy and compliance. *Organic Agriculture* 5:53-62.
- Wan, J., Guo, J., Miao, Z., and Guo, X., 2016. Reverse micellar extraction of bromelain from pineapple peel effect of surfactant structure. *Food Chemistry* 197:450-456.
- Wang, B., Li, R., Ruan, Y., Ou, Y., Zhao, Y., and Shen, Q., 2015. Pineapple-banana rotation reduced the amount of Fusarium oxysporum more than maize-banana rotation mainly through modulating fungal communities. *Soil Biology & Biochemistry* 86:77-86.
- Wang, L., Tang, D., Kuang, Y., Lin, F., and Su, Y., 2015. Structural characteristics of pineapple pulp polysaccharides and their antitumor cell proliferation activities. *Journal of the Science of Food and Agriculture* 95:2554-2561.
- Wittayakun, S., Innaree, S., Innaree, W., and Chainetr, W., 2015. Supplement of sodium bicarbonate, calcium carbonate and rice straw in lactating dairy cows fed pineapple peel as main roughage. *Slovak Journal of Animal Science* 48:71-78.

- Wong, C.W., Pui, L.P., and Ng, J.M.L., 2015. Production of spray-dried Sarawak pineapple (*Ananas comosus*) powder from enzyme liquefied puree. *International Food Research Journal* 22:1631-1636.
- Yeboah, N.E., Feng, Y., and Evelyn, E., 2015. Embracing risk mitigation strategies in pineapple supply chain and its impact on supply chains performance. *Asian Journal of Agricultural Extension, Economics and Sociology* 4:302-316.
- Youryon, P. and Wongs-Aree, C., 2015. Postharvest application of calcium chloride affects internal browning reduction during low temperature storage of 'Sawi' pineapple. *Acta Horticulturae* 1088:197-200.
- Yuris, A. and Siow, L.-F., 2014. A Comparative Study of the Antioxidant Properties of Three Pineapple (*Ananas comosus L.*) Varieties. *Journal of Food Studies* 3.
- Zhang, Q., Liu, Y., He, C., and Zhu, S., 2015. Postharvest exogenous application of abscisic acid reduces internal browning in pineapple. *Journal of Agricultural and Food Chemistry* 63:5313-5320.
- Zhou, L., Matsumoto, T., Tan, H., Meinhardt, L.W., Mischke, S., Wang, B., and Zhang, D., 2015. Developing single nucleotide polymorphism markers for the identification of pineapple (*Ananas comosus*) germplasm. *Horticulture Research 2,Article number:* 15056 (2015) 2.

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